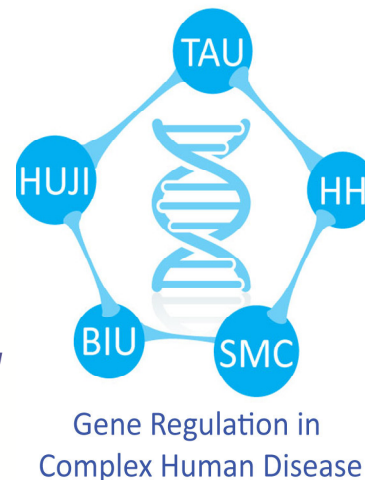


Chromatin States: A quantitative genetics perspective

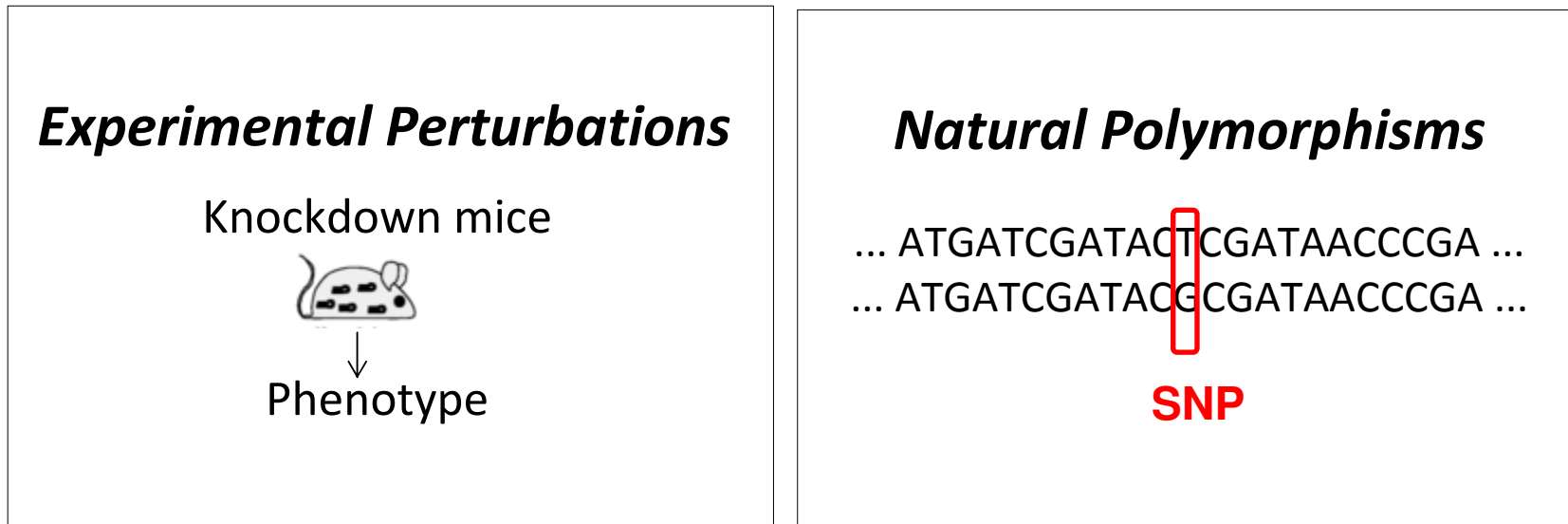
Irit Gat-Viks

Department of Cell research and Immunology
Life Sciences, Tel-Aviv University



The genetic basis of disease

Finding genes influencing a disease



Exploit natural polymorphisms to understand disease mechanisms

Genetic variant



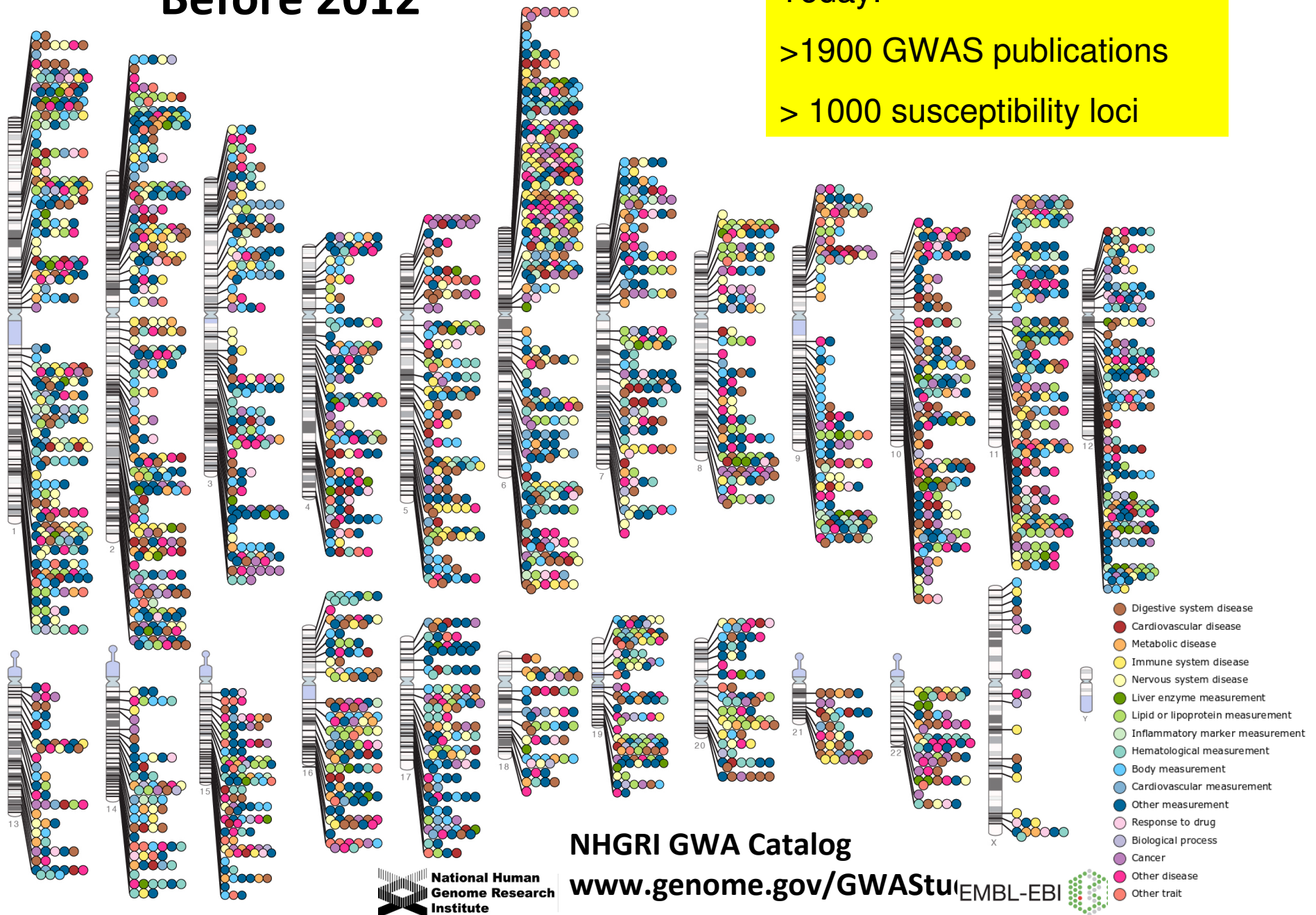
Genome-wide association studies (GWAS)

Complex phenotype



Before 2012

Today:
>1900 GWAS publications
> 1000 susceptibility loci

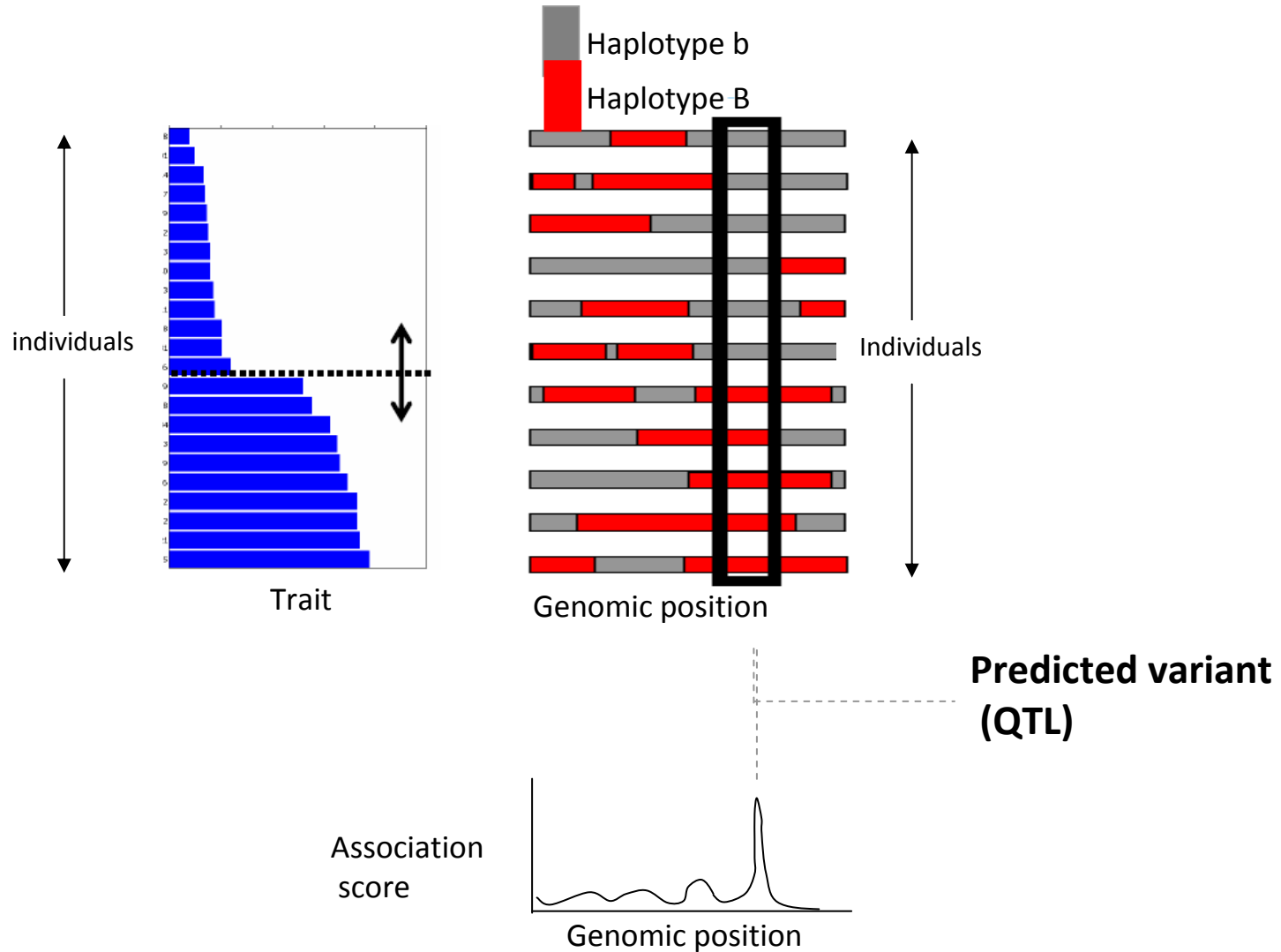


NHGRI GWA Catalog

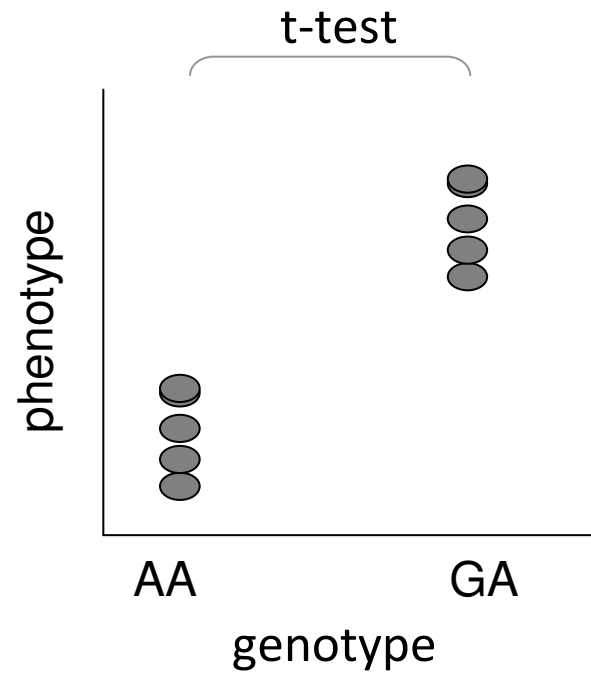
www.genome.gov/GWASu EMBL-EBI



Association study for a quantitative trait



QTL analysis: Standard linear models



QTL analysis: Standard linear models

1. ANOVA model

$$x_{ij} = \mu_i + e_{ij}$$

x_{ij} - Phenotype of j-th individual of marker genotype i

μ_i - Effect of marker i

e_{ij} - Residual error: the deviation of the jth individual from the expected value of the ith marker, $E(e_{ij})=0$. $\text{Var}(e_{ij})=\sigma^2$

The presence of a linked QTL is indicated by a significant between-marker variance

2. Multiple regression model

$$x_j = \mu + \sum_{i=1}^n b_i g_{ij} + e_j$$

x_j - Phenotype of jth individual

g_{ij} - Indicator variables (one for each marker genotype)

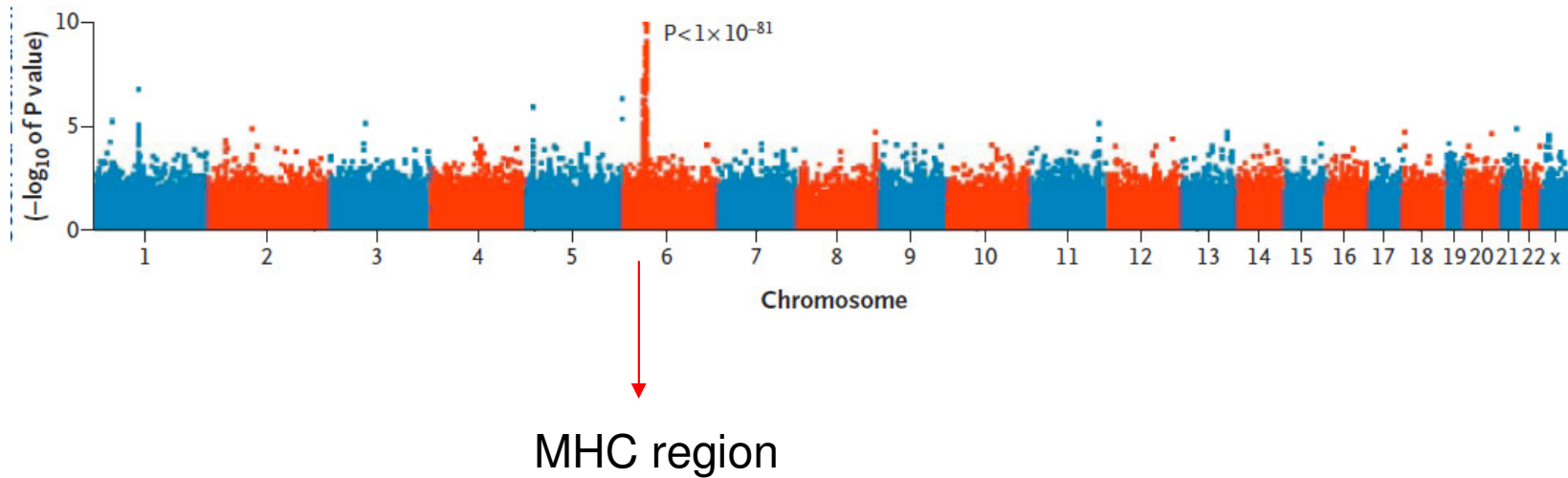
$$g_{ij} = \begin{cases} 1 & \text{if individual } j \text{ has marker genotype } i \\ 0 & \text{otherwise} \end{cases}$$

e_j - Residual error

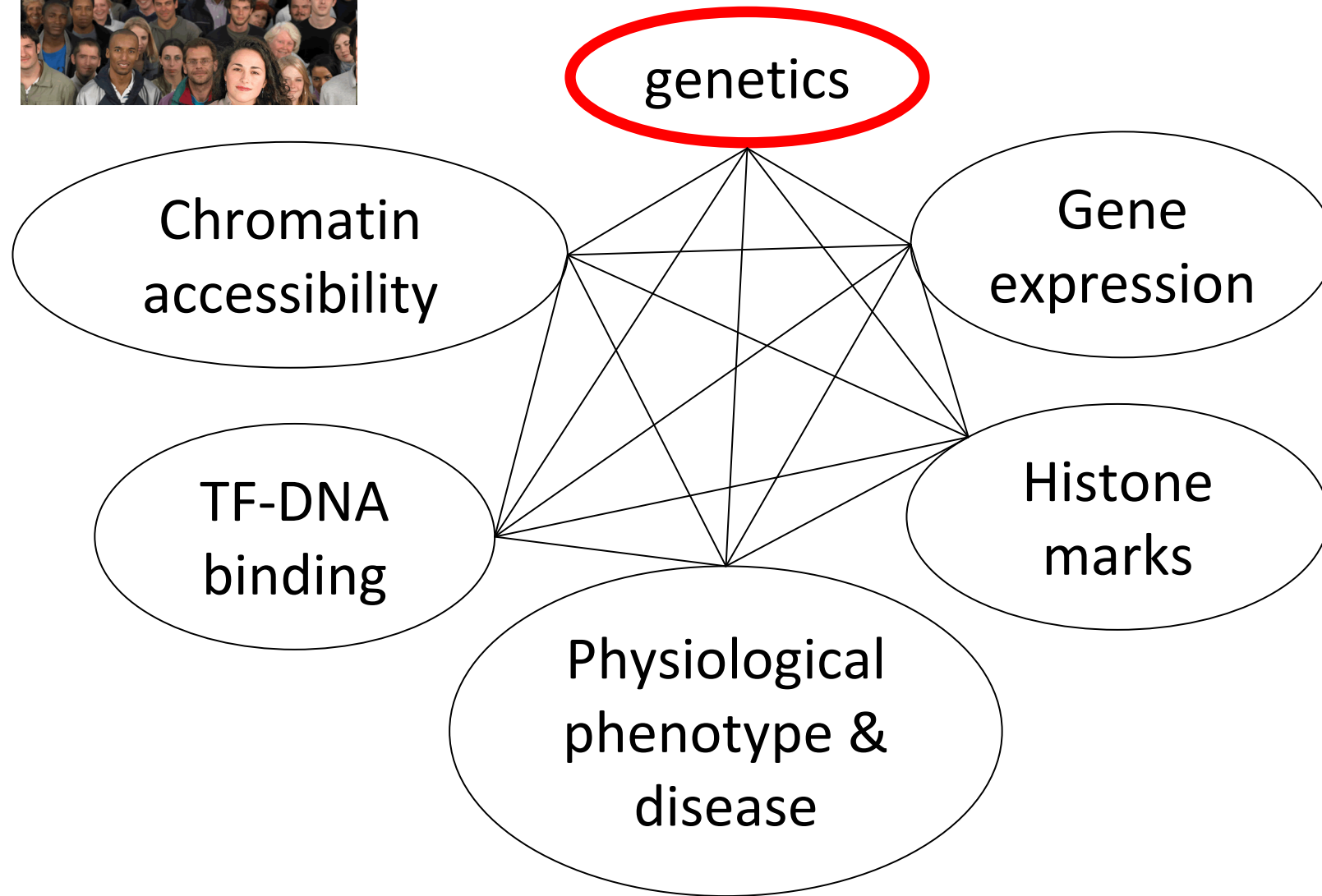
The presence of a linked QTL is indicated by a significant fraction of character variance accounted for by the marker genotype

Risk Alleles for Multiple Sclerosis

Manhattan plot



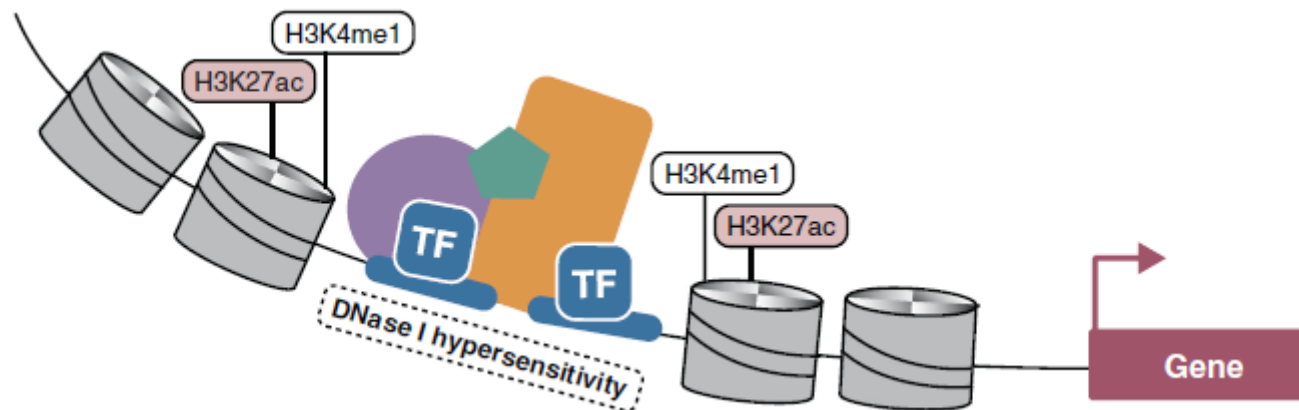
A quantitative genetics perspective



Constructing **regulatory elements maps** using epigenomic profiling

For example, revealing **enhancers**:

- The locations of enhancer elements coincide with DNase I hypersensitive regions of open chromatin flanked by nucleosomes marked with H3K4me1/2.
- H3K27ac and H4K16ac are associated with active chromatin.
- H3K27me3 and H3K9me3 are associated with repressed chromatin.



Interpreting susceptibility loci using epigenomic profiling

Phenotyping + Genotyping

↓
GWAS

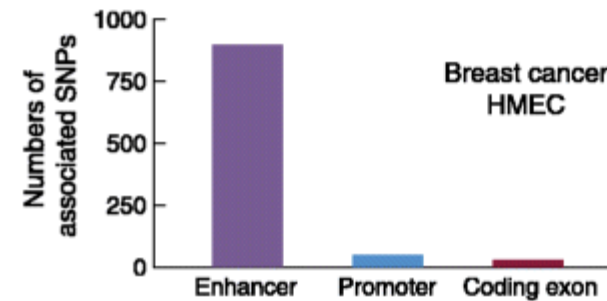
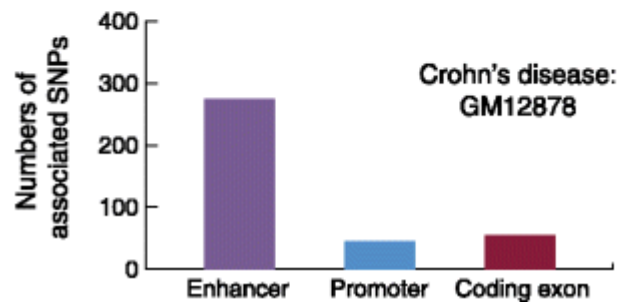
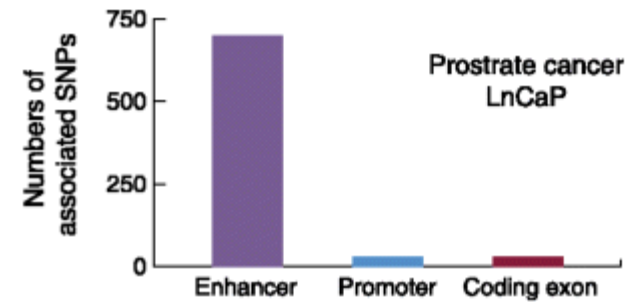
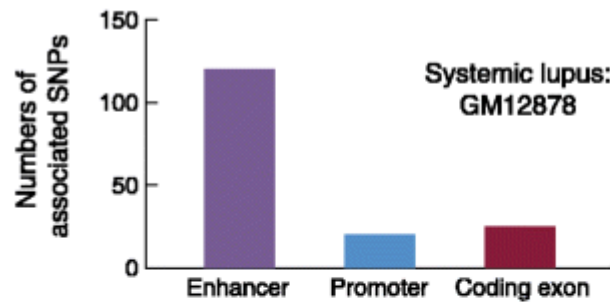
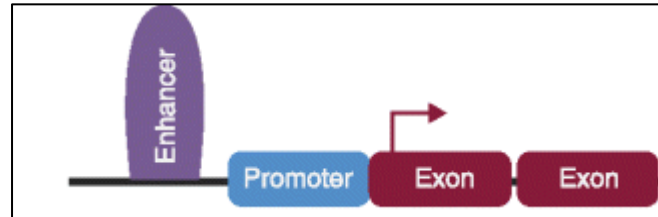
↓
Genetic risk variants

Epigenomic profiling

↓
Regulatory elements maps

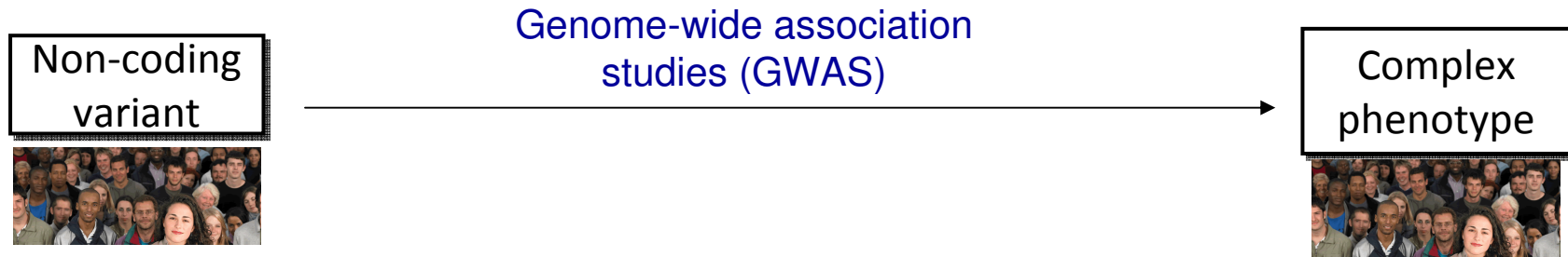
- Loci with enhancer features (H3K4me1, H3K27ac) are highly enriched for risk variants
- Risk variants preferentially map to enhancers specific to disease-relevant cell types (e.g., colon cancer predisposition variants)

Enrichment of genome-wide association study variants in putative enhancer elements

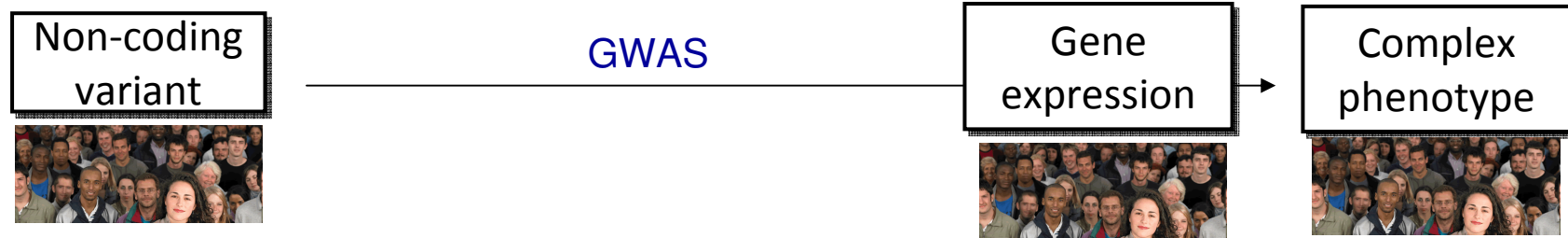


The challenge: what is the **role** of non-coding variants in gene expression?

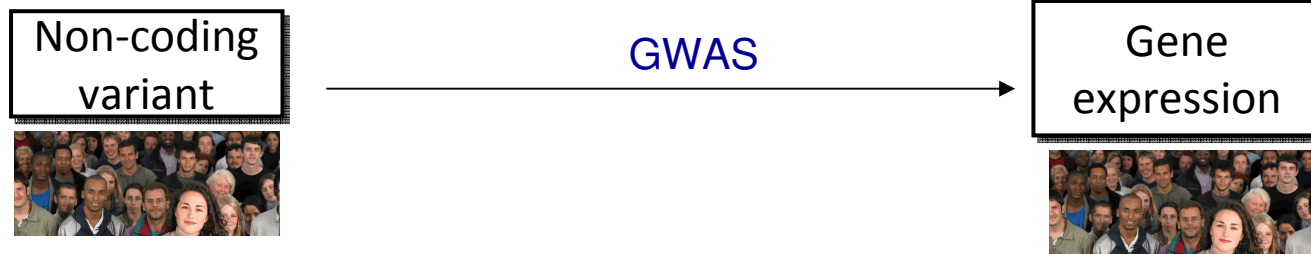
Understand the role of non-coding variants in gene expression



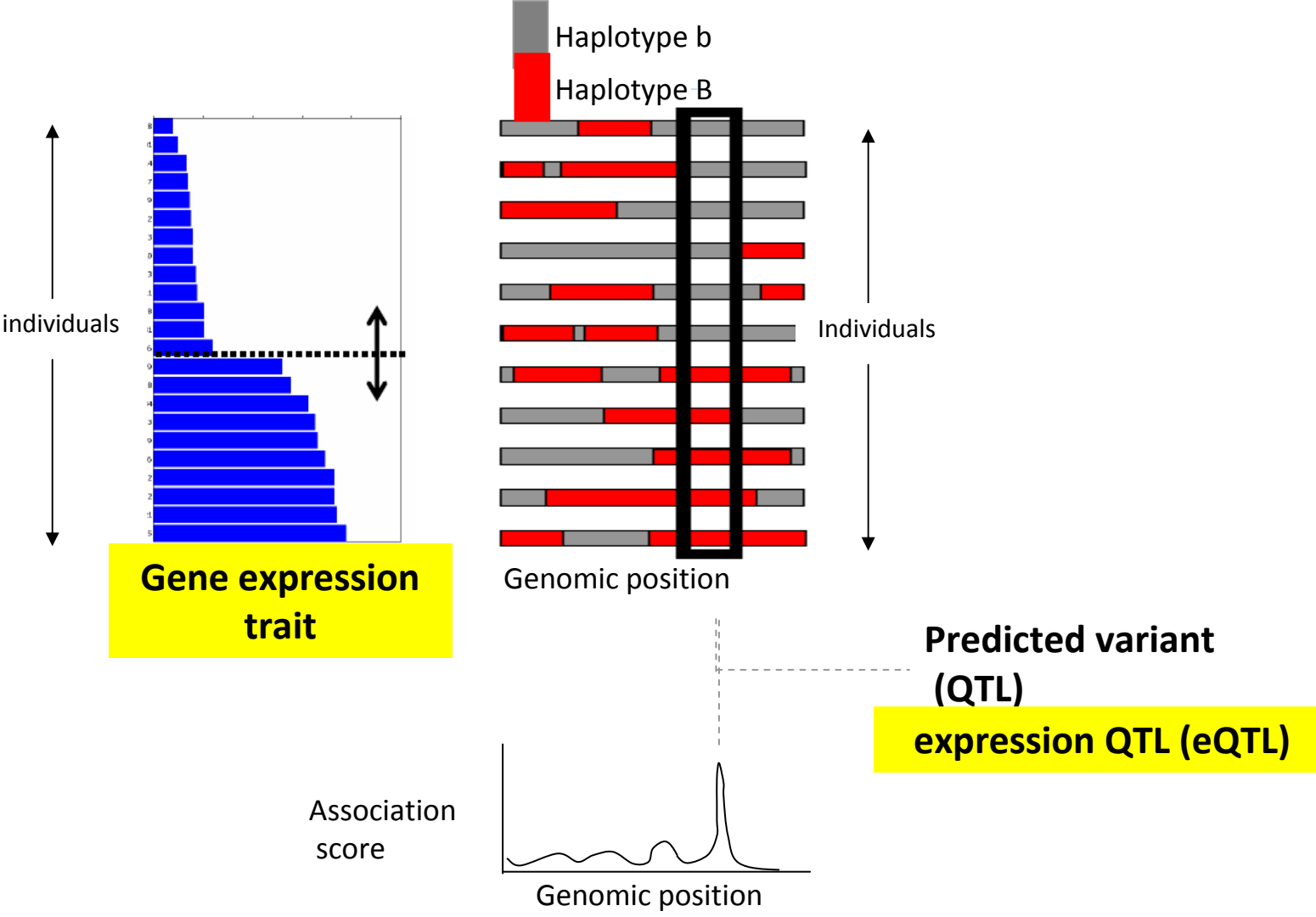
Understand the role of non-coding variants in gene expression



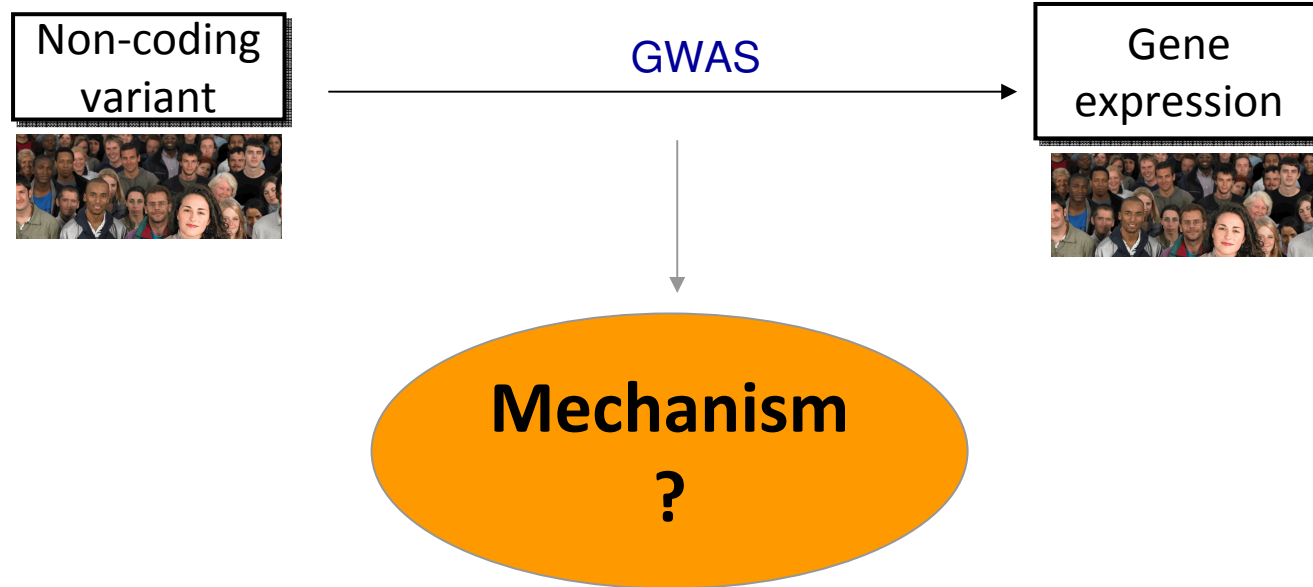
Understand the role of non-coding variants in gene expression



Understand the role of non-coding variants in gene expression

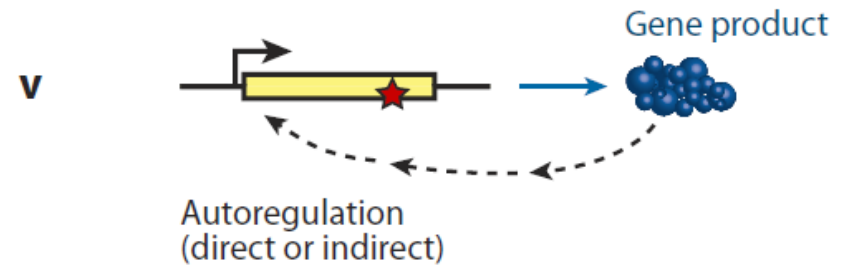
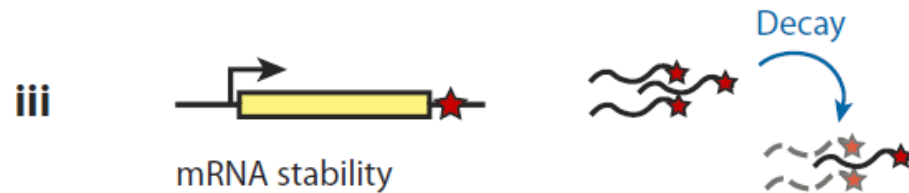
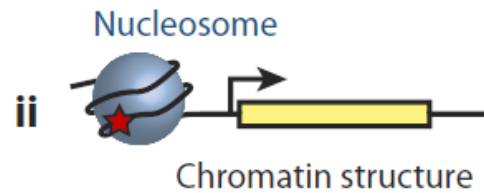
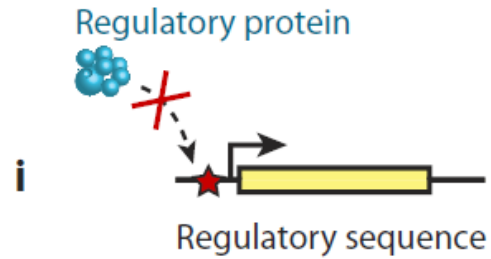


Understand the role of non-coding variants in gene expression



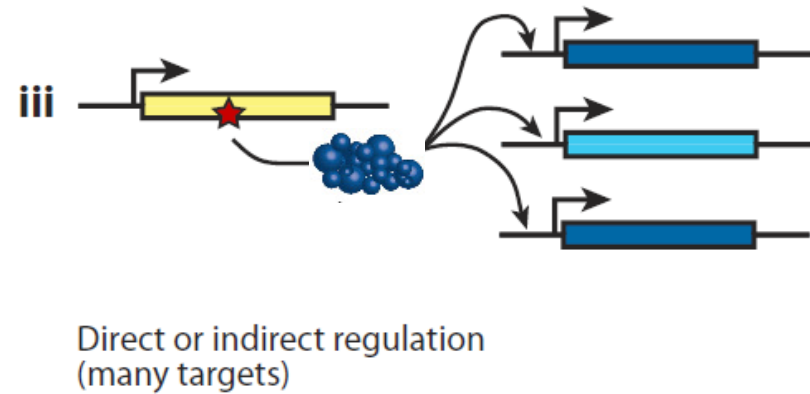
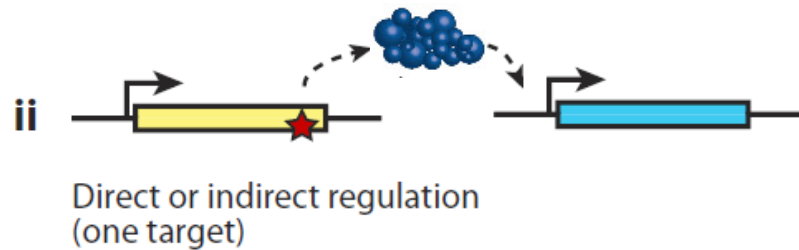
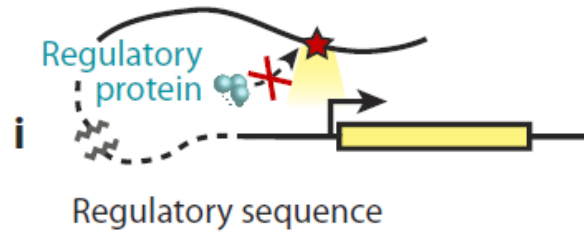
Understanding the role of variants in gene expression based on their genomic positions

a Local



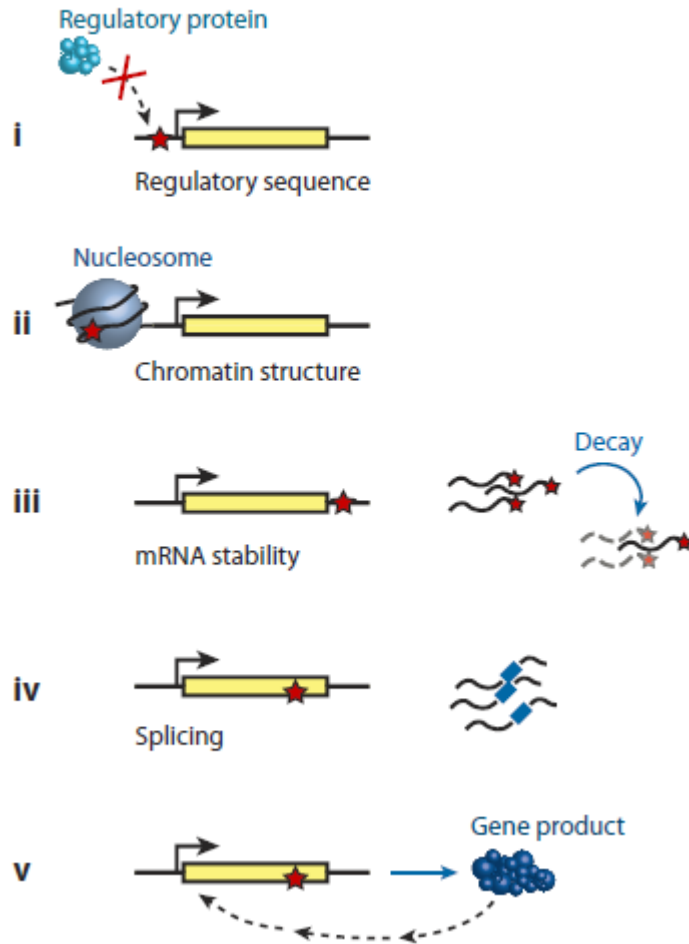
Understanding the role of variants in gene expression based on their genomic positions

b Distant



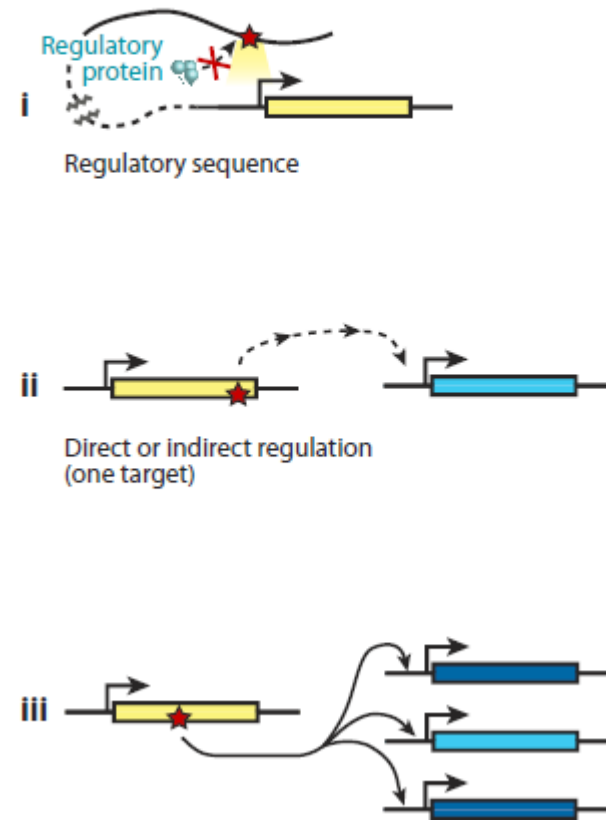
Understanding the role of variants in gene expression based on their genomic positions

a Local



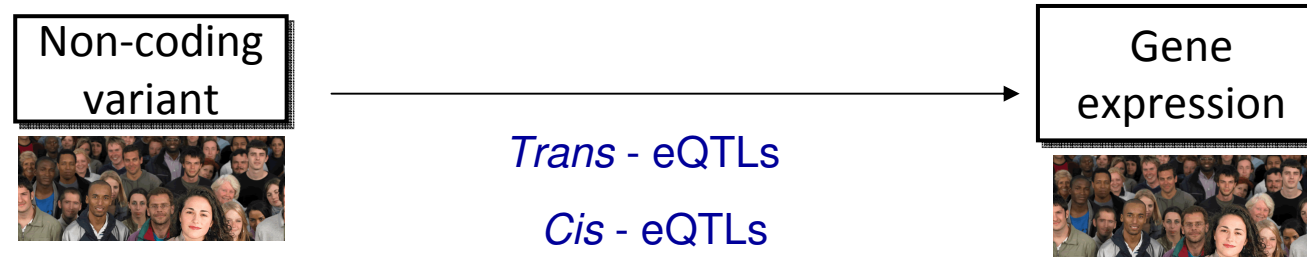
Cis-acting eQTL

b Distant



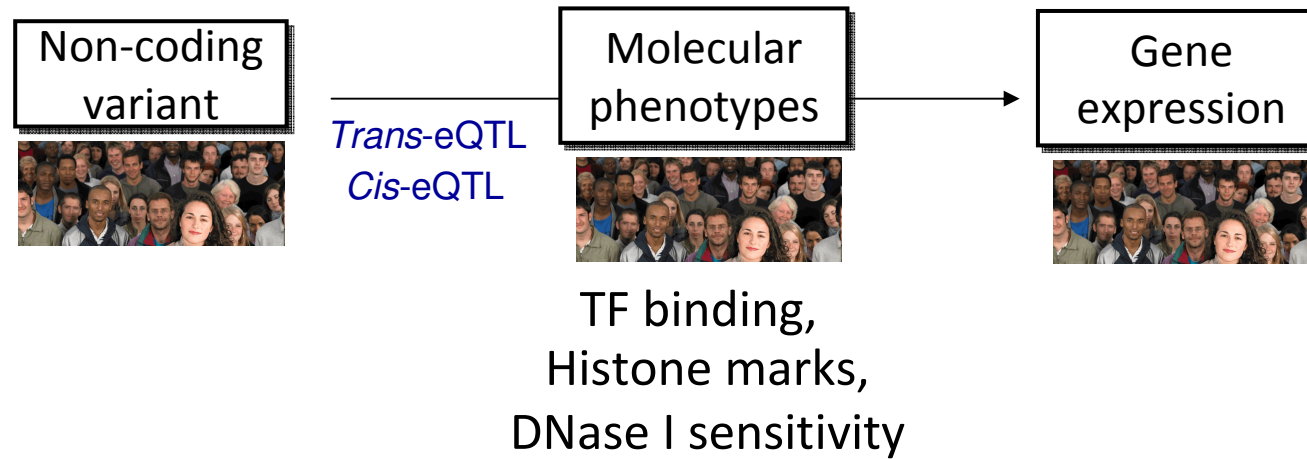
Trans-acting eQTL

Determine the mechanisms by which regulatory variants affect gene expression

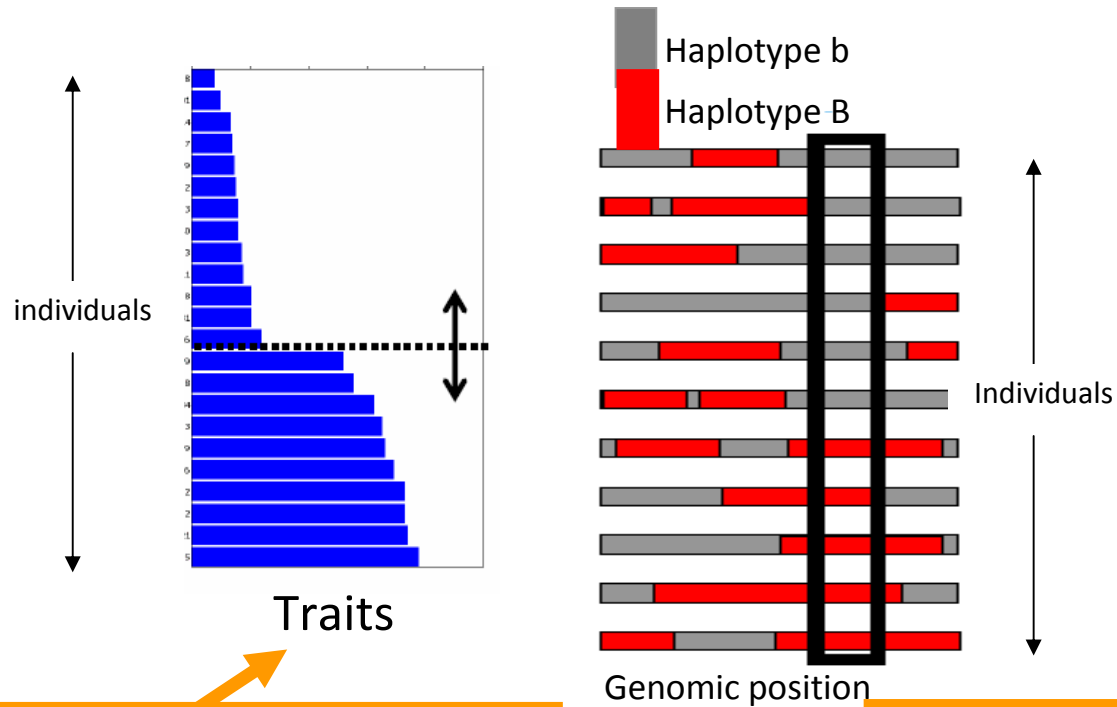


The challenge: eQTL analysis cannot reveal the complete functional mechanism by which non-coding variants influence gene expression

Determine the mechanisms by which regulatory variants affect gene expression



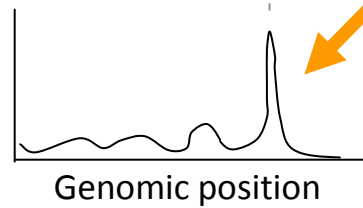
Association study for a quantitative trait



- Gene expression traits
- Open chromatin traits
- Histone modification traits

- *Cis*-association
- *Trans*-association

Association score



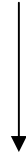
Example 1: Genetic landscape of open chromatin in yeast

Example 1: Genetic landscape of open chromatin in yeast

FAIRE-seq



Identify open chromatin peaks (OCRs)

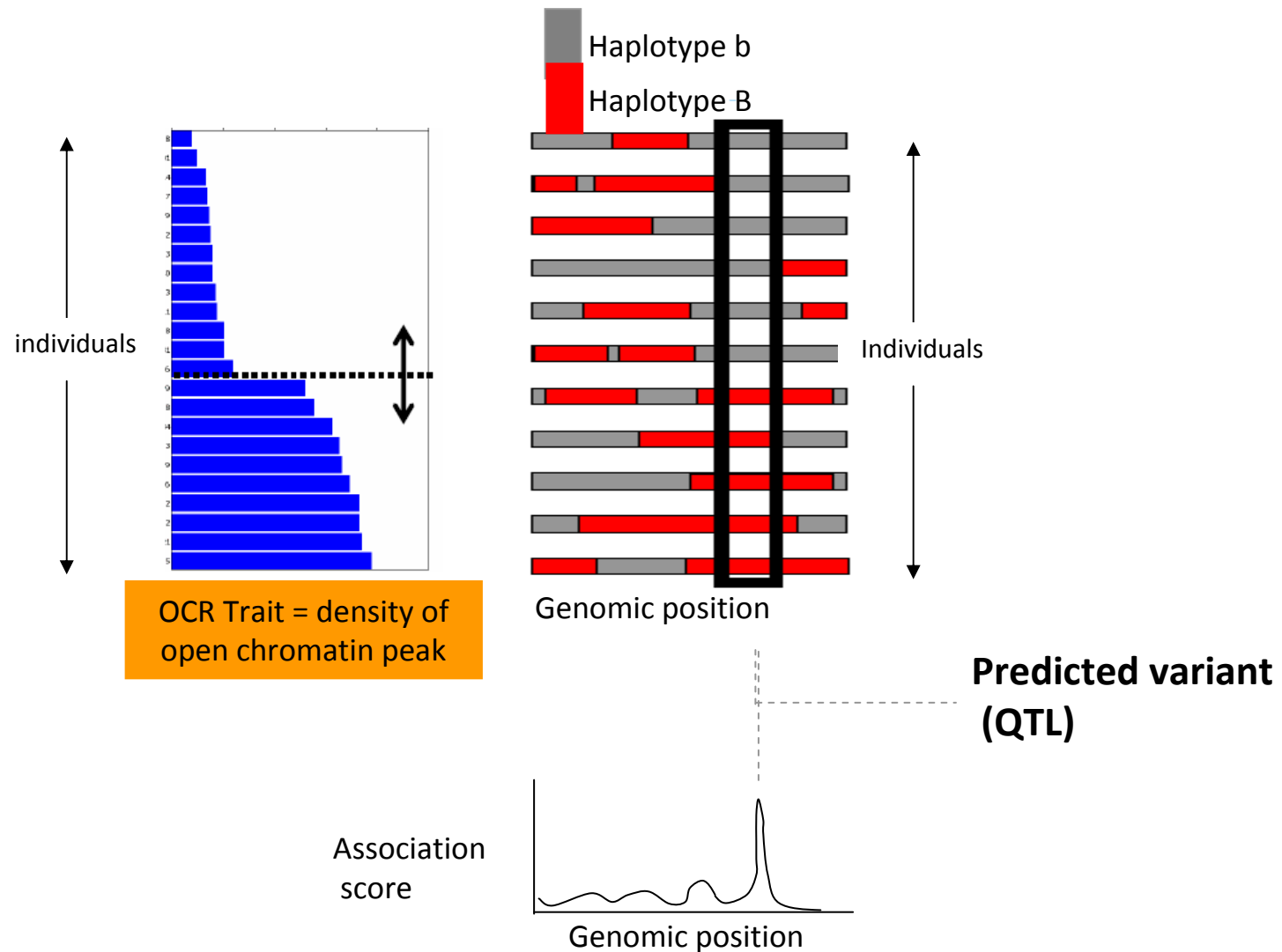


For each peak: **QTL analysis** of OCR traits

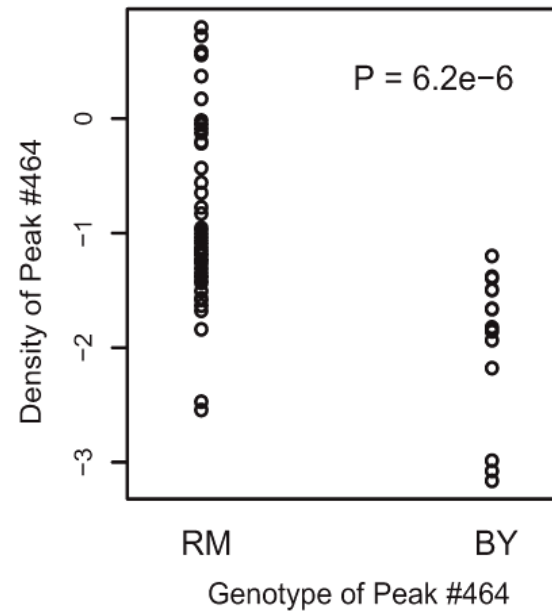
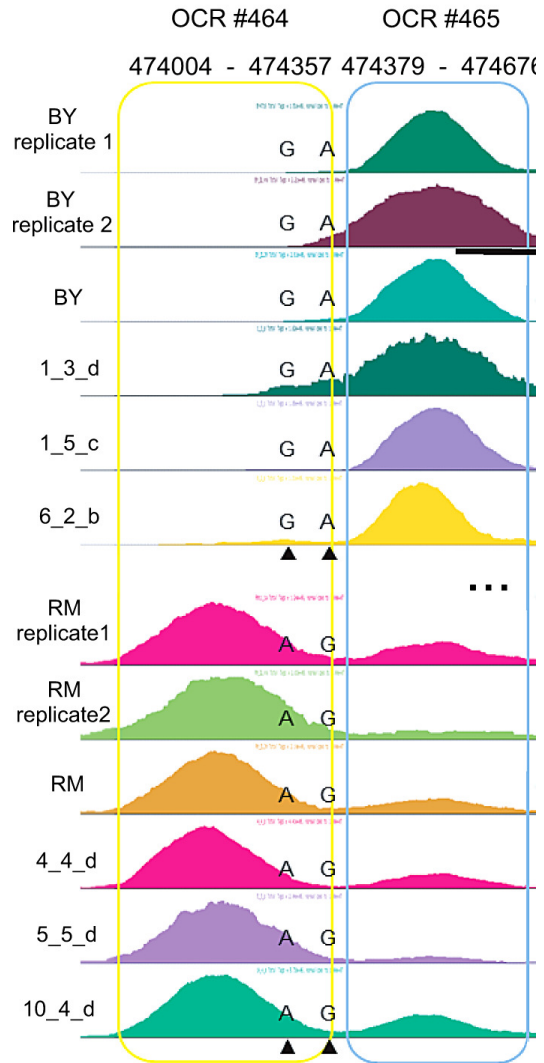
OCR Trait = density of peak

Broader peaks of naked
DNA compared to DHS-seq
and ATAC-seq

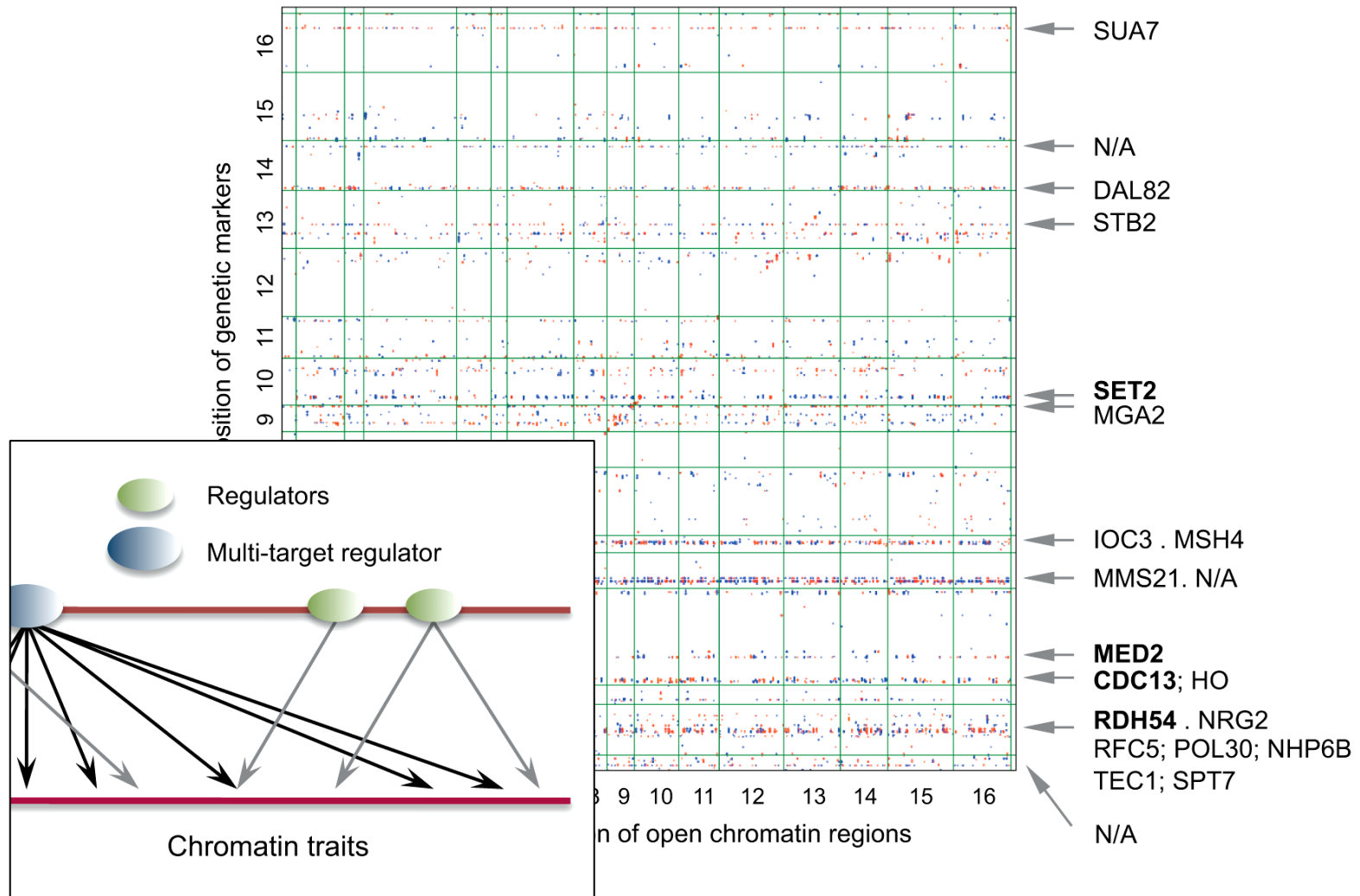
Association study for an open chromatin trait



Characterization of *cis*-associations



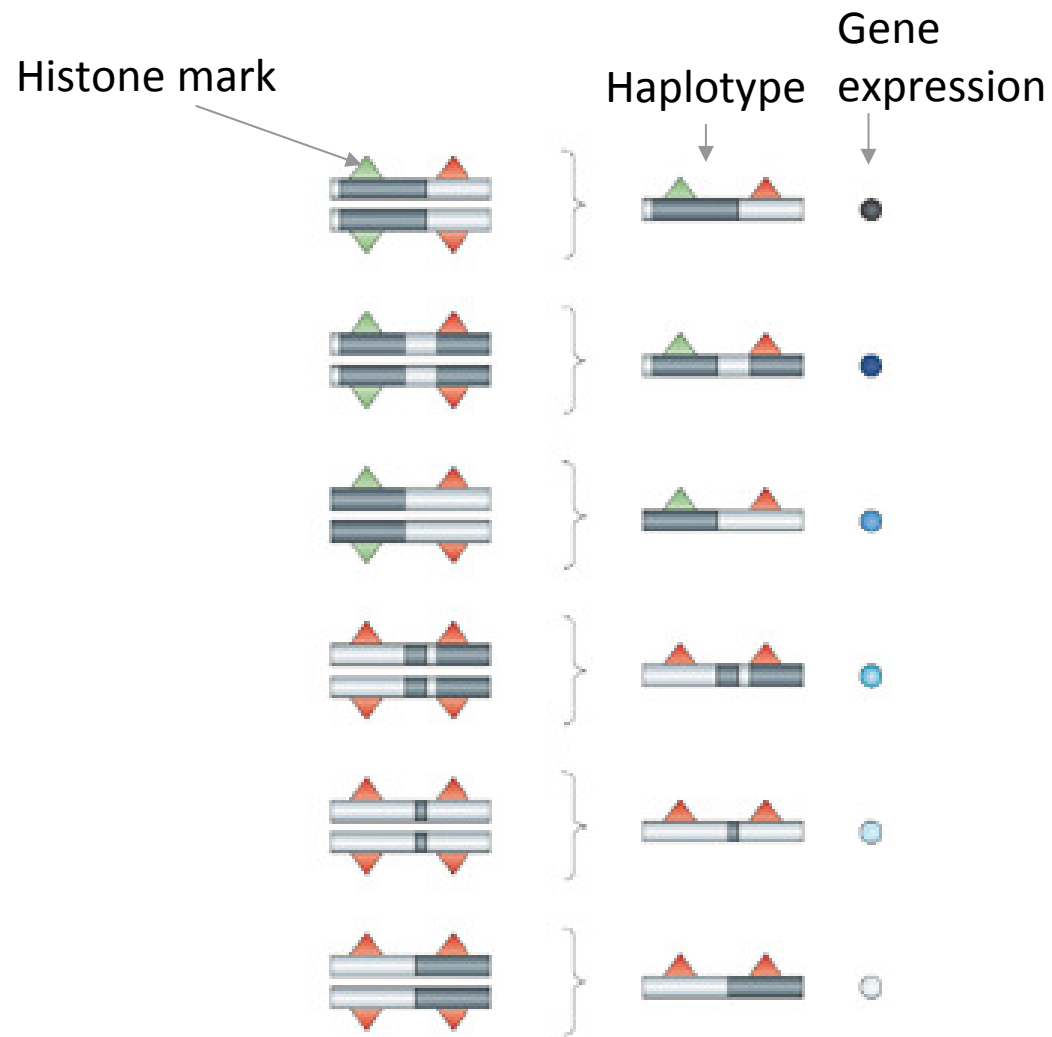
Characterization of *trans*-associations



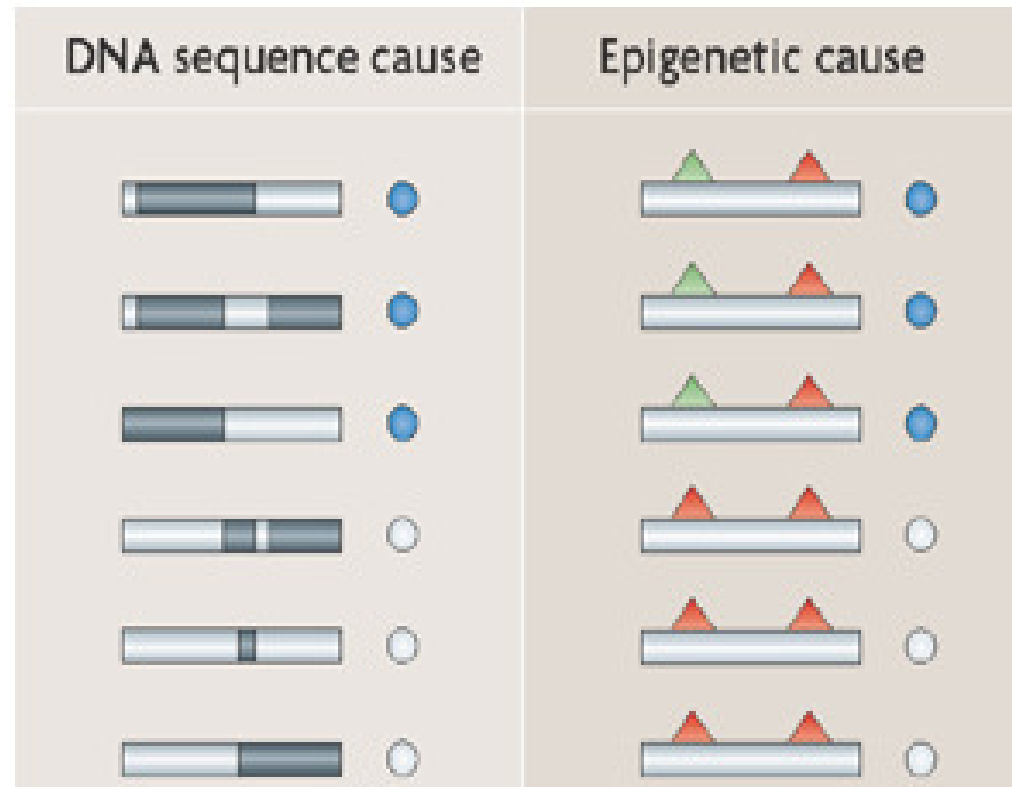
Example 2:

Integrate genetics + histone marks + expression traits

Integrate genetics + histone marks + expression traits



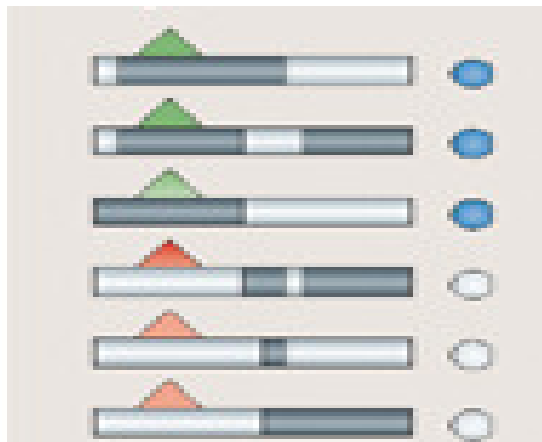
Integrate genetics + histone marks + expression traits



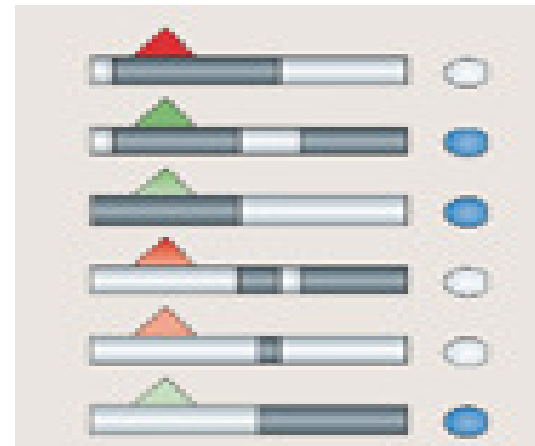
Integrate genetics + histone marks + expression traits

Relationships between
histone marks and gene
expression

Sequence
dependent

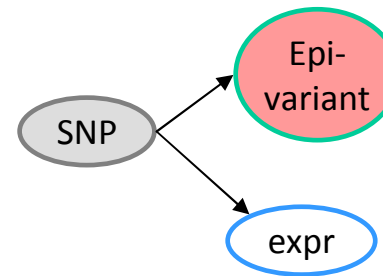
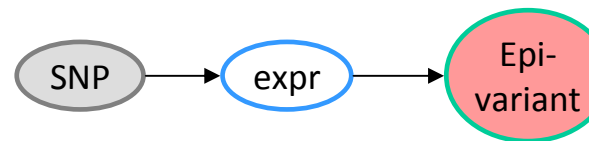
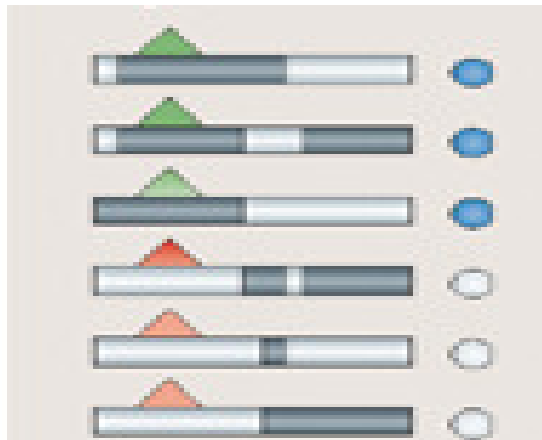


Sequence
independent

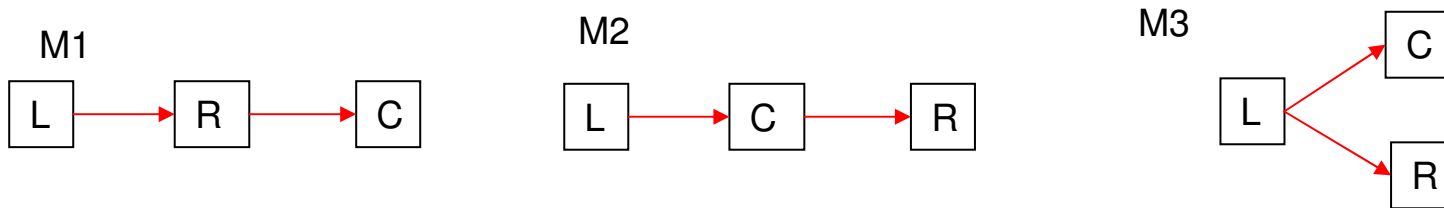
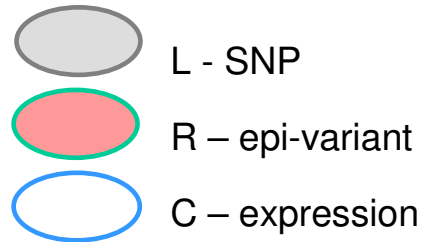


Integrate genetics + histone marks + expression traits

Sequence dependent



Inferring causal relations




$$M1. P(L, R, C) = P(L) P(R|L) P(C|R)$$

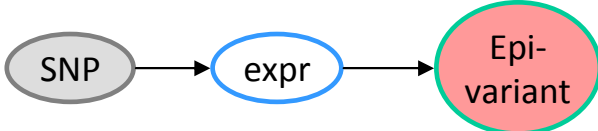
$$M2. P(L, R, C) = P(L) P(C|L) P(R|C)$$

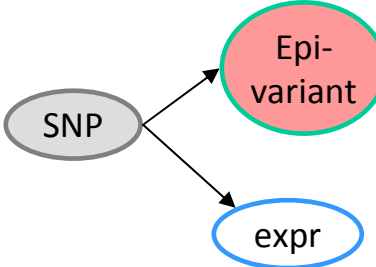
$$M3. P(L, R, C) = P(L) P(C|L) P(R|C, L)$$

The likelihood for each model over all individuals in the population of interest are given by:

$$\text{likelihood function} = L(\theta; M) = p(\text{data} | \theta_M)$$

$$P(L, R, C | \theta_{M1}) = \prod_{i=1}^n P(L_i) \cdot P(R_i | L_i) \cdot P(C_i | R_i)$$


$$P(L, R, C | \theta_{M2}) = \prod_{i=1}^n P(L_i) \cdot P(C_i | L_i) \cdot P(R_i | C_i)$$


$$P(L, R, C | \theta_{M1}) = \prod_{i=1}^n P(L_i) \cdot P(R_i | L_i) \cdot P(C_i | L_i)$$


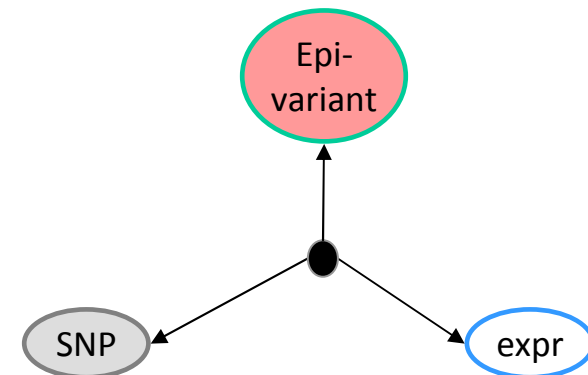
Integrate genetics + histone marks + expression traits

Rintish et al. 2014; 30 Rat BXH/BXB strains, liver and heart

- 18.1% and 14.5% of all H3K4me3 and H3K27me3 QTLs were also eQTL.
- 20% of all eQTL were also QTL for a histone mark.

Degner et al. (2012), 70 Yoruba lymphoblastoid cell lines

- 16% of DNase I sensitivity QTL (dsQTL) were also eQTL
- 23% of eQTL were also dsQTL



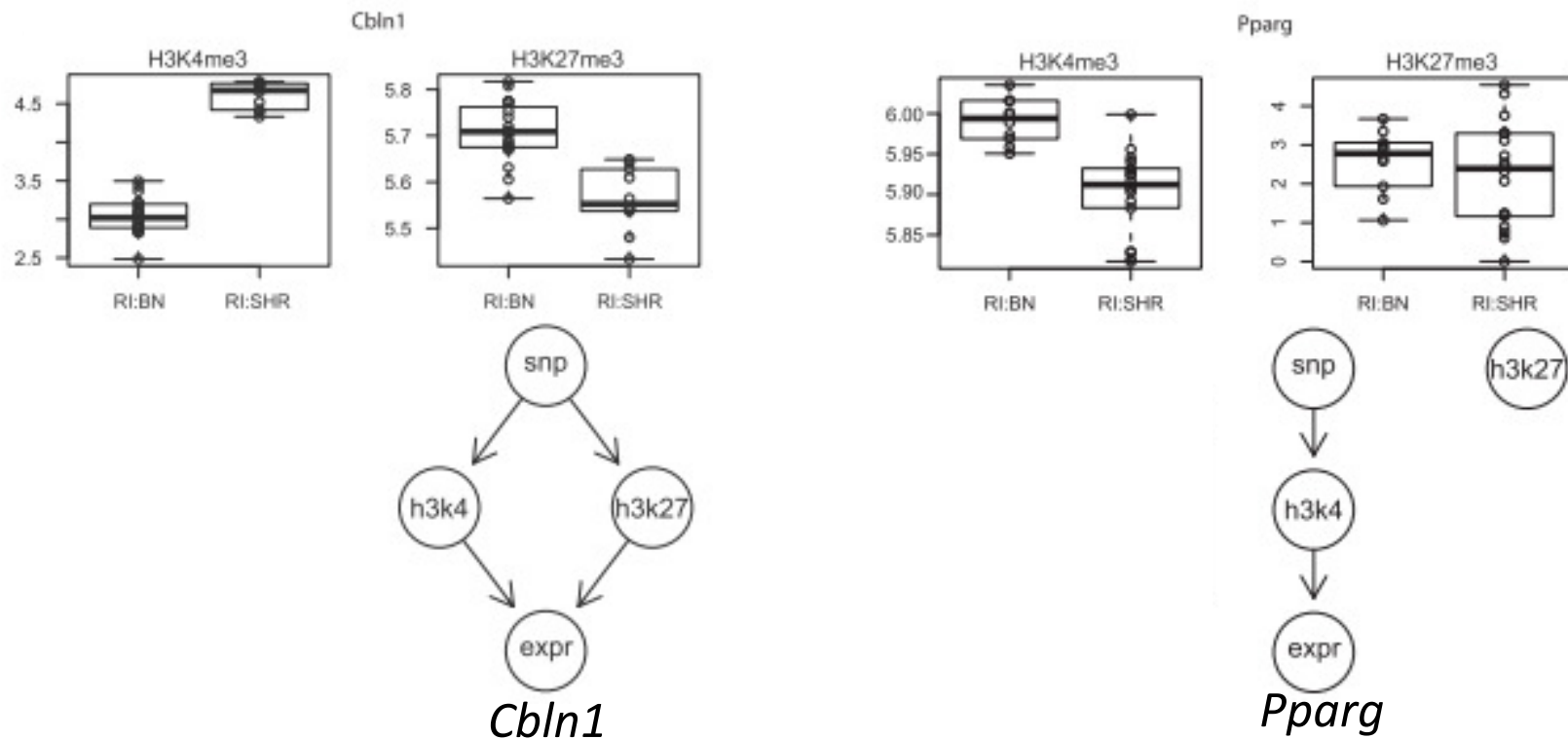
Genetic landscape of histone modifications in rat liver and heart

Table 2. HistoneQTL mapping results in heart and liver tissue

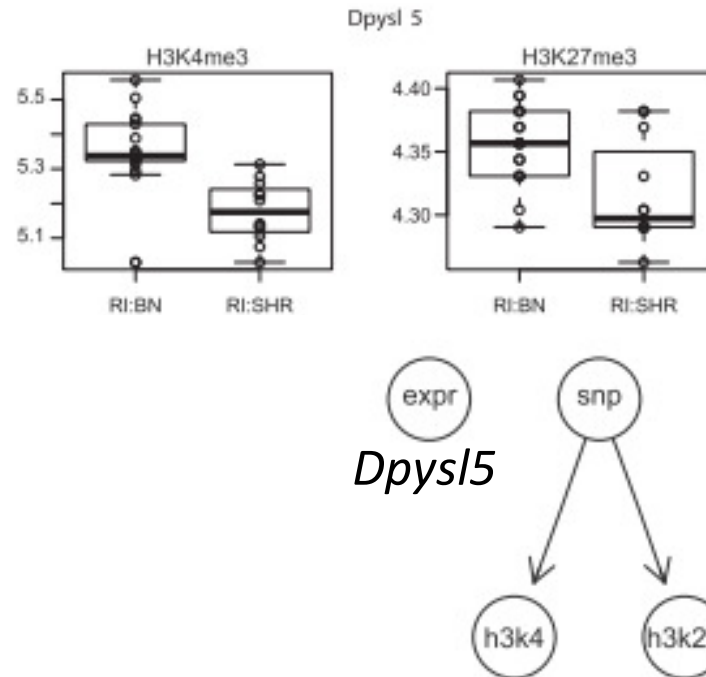
	Heart		Liver		Both tissues	
H3K4me3 (traits)	(25,064)		(31,447)		(20,076)	
FDR-cutoff ^a	<i>cis</i>	<i>trans</i>	<i>cis</i>	<i>trans</i>	<i>cis</i>	<i>trans</i>
0.05	2638	2504	2945	1464	931	82
0.01	2024	812	2235	243	698	31
0.001	1414	232	1454	59	467	13
1×10^{-4}	776	30	834	26	232	4
1×10^{-5}	0	0	460	12	0	0
H3K27me3 (traits)	(4,214)		(3,776)		(2,688)	
FDR-cutoff ^a	<i>cis</i>	<i>trans</i>	<i>cis</i>	<i>trans</i>	<i>cis</i>	<i>trans</i>
0.05	102	15	196	38	35	1
0.01	74	4	166	7	30	1
0.001	57	2	131	2	23	0
1×10^{-4}	50	1	97	0	18	0
1×10^{-5}	50	1	97	0	18	0

^aCutoff for limitation of false discoveries.

Integrate genetics + histone marks + expression traits



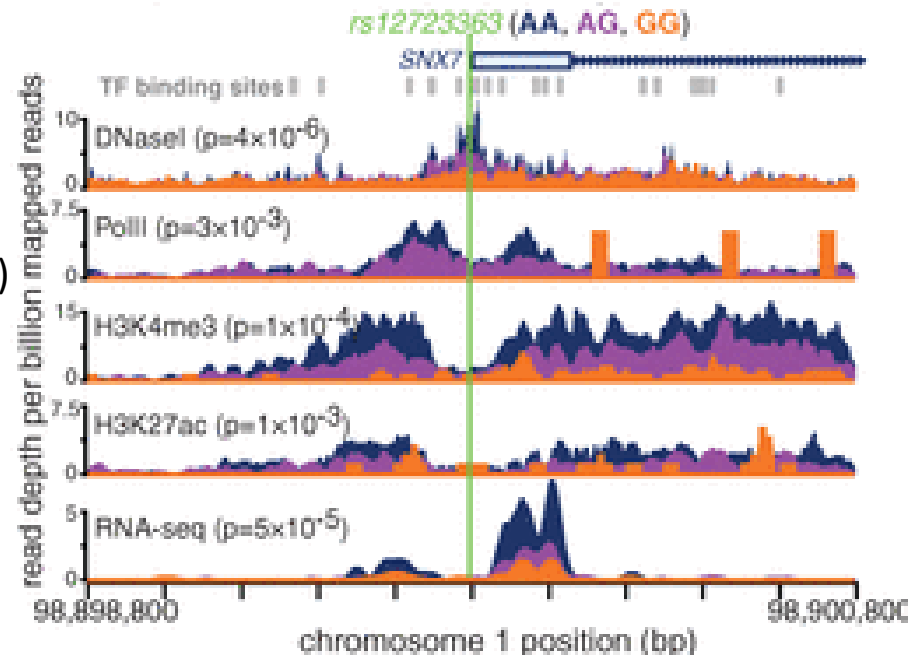
Integrate genetics + histone marks + expression traits



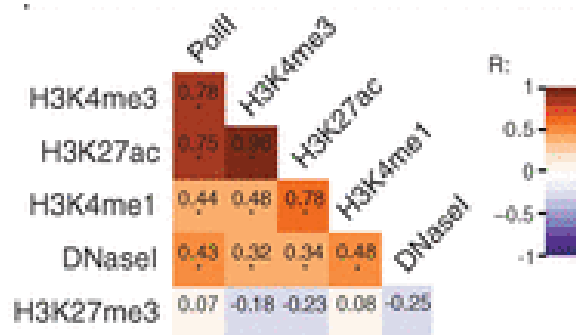
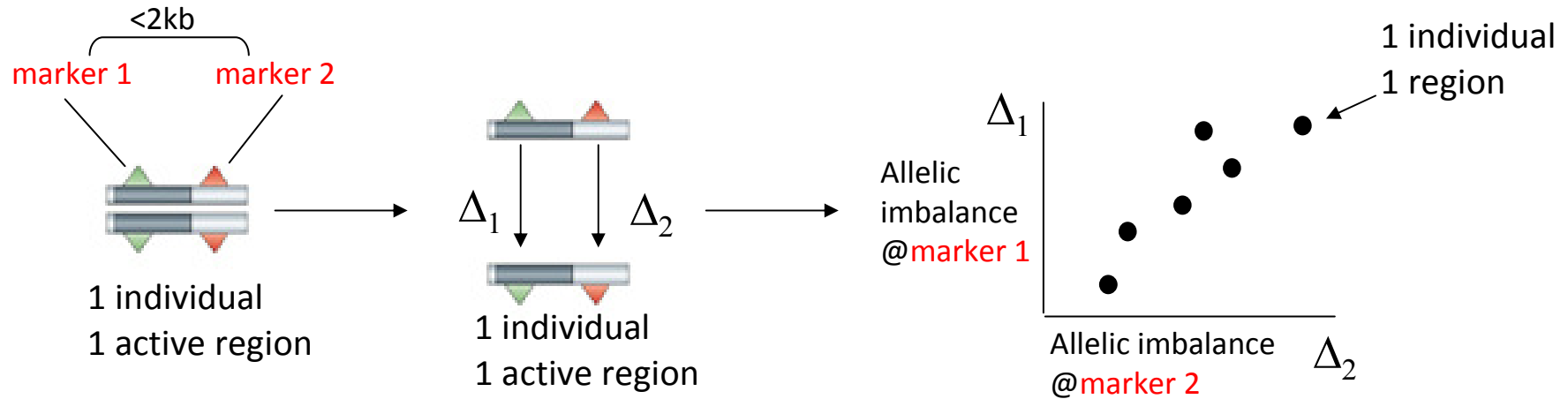
Example 3: Revealing functionally linked modifications

Coordinated change in histone marks along ~2kb regions

Lymphoblastoid cell lines (LCLs)
from 70 Yoruba (Nigeria)
individuals

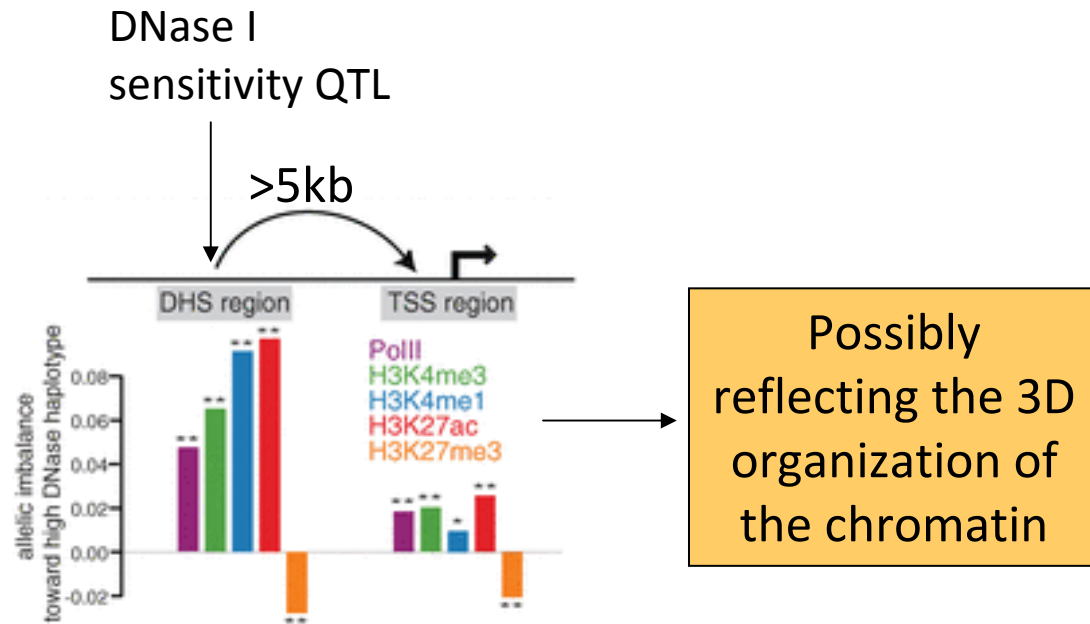


Revealing functionally linked modifications, depending on the same genetic element



Correlation in **allelic imbalance** between histone marks at DNase I sensitive QTL sites (dsQTLs)

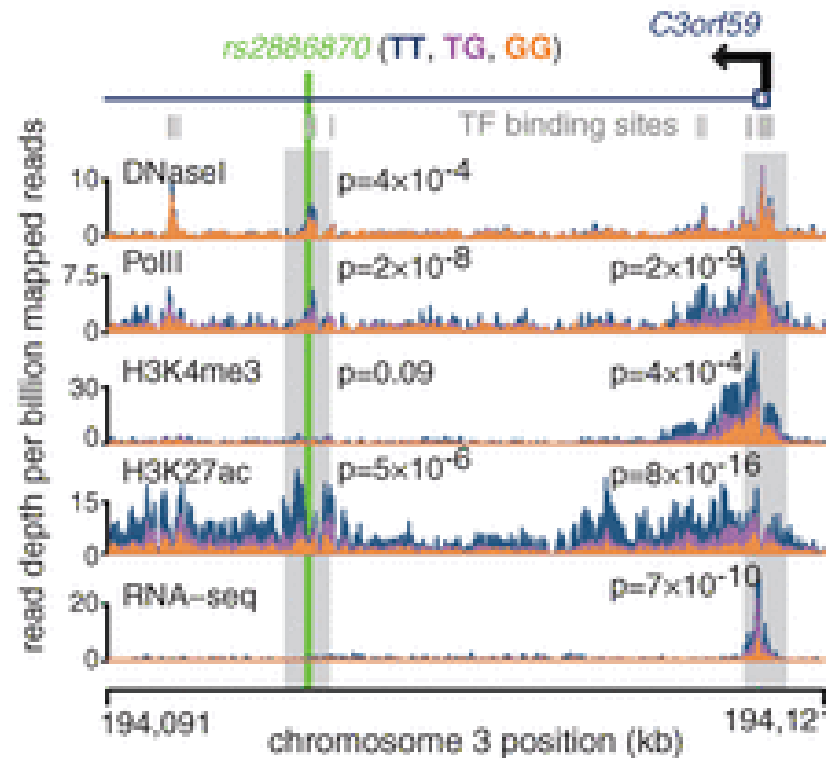
Coordinated change in histone marks between distal (>5kb) regions



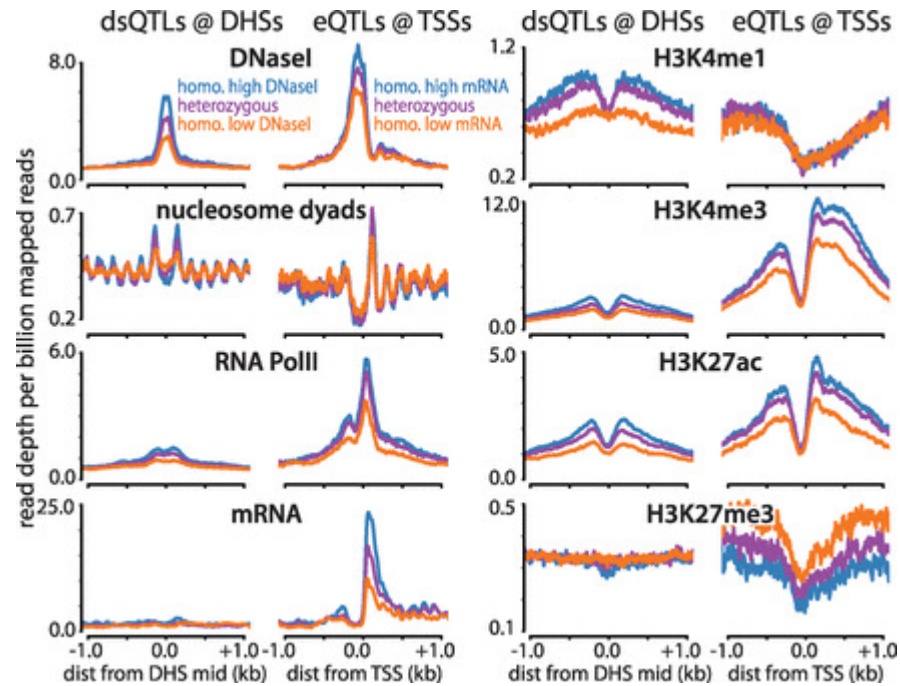
Allelic imbalance across DNase I sensitivity QTLs and eQTLs

ChIP-Seq mark	Functional association
H3K4me3	Active promoters
H3K4me1	Active enhancers
H3K27ac	Active promoters and enhancers
H3K27me3	Inactive chromatin
RNA Pol II	Transcription

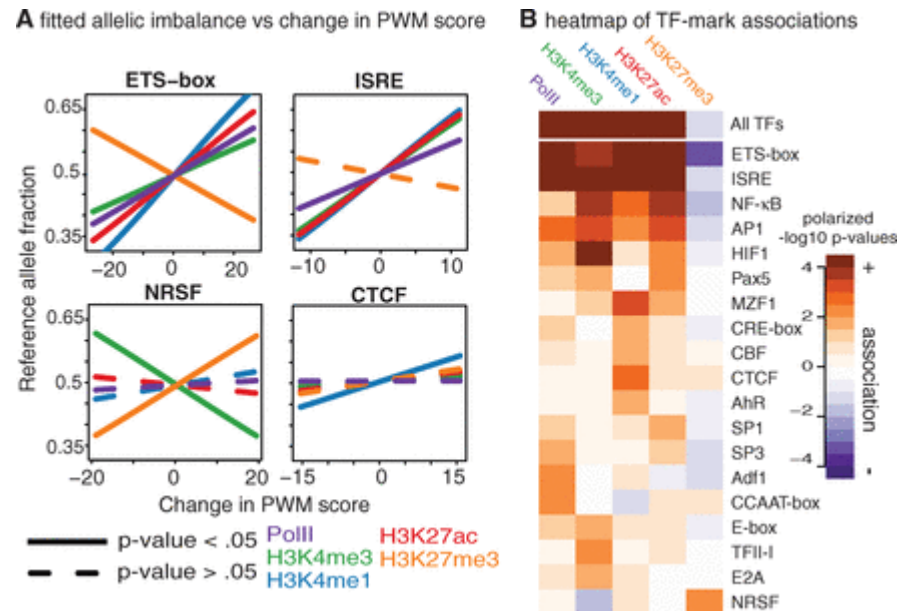
Coordinated change in histone marks between distal (>5kb) regions



Coordinated change in histone marks between distal regions

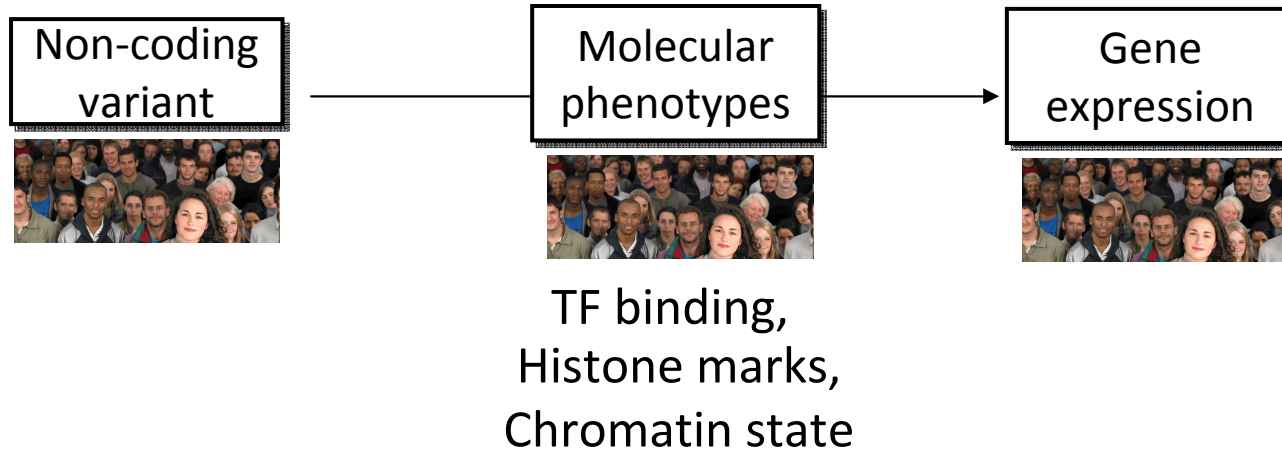


Coordinated change in histone marks between distal regions

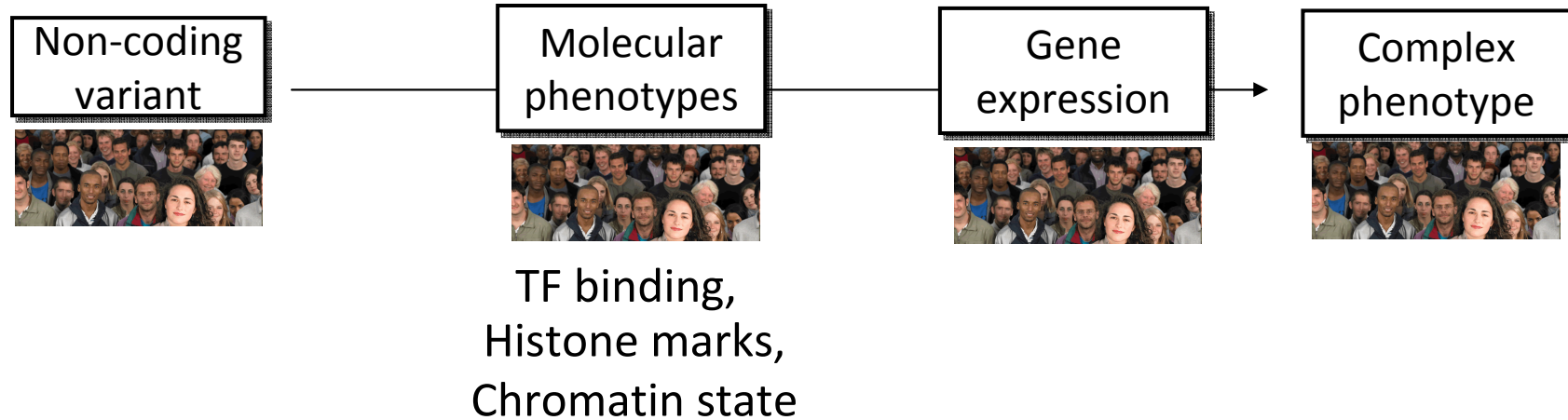


Example 4: Reveal the function of chromatin states
in complex physiological traits

Example 4: Reveal the function of chromatin states in complex physiological traits



Example 4: Reveal the function of chromatin states in complex physiological traits

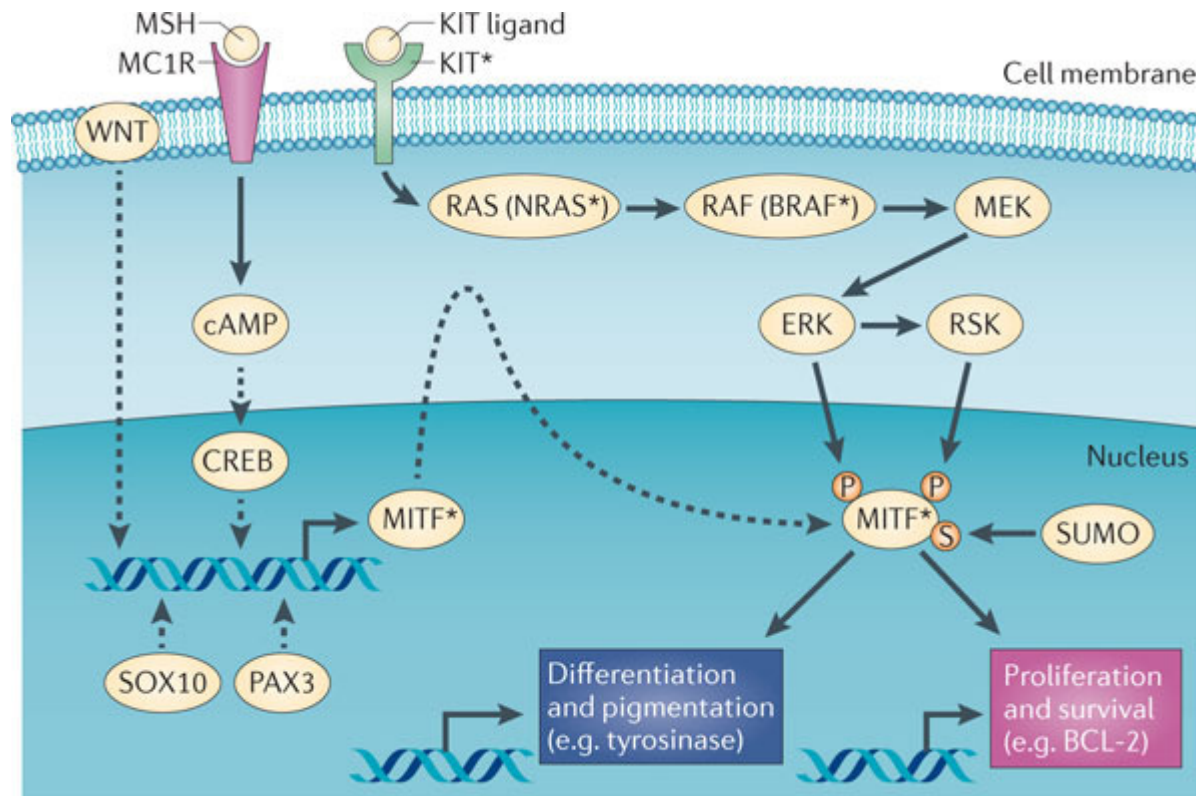


A molecular basis for classic blond hair color in Europeans

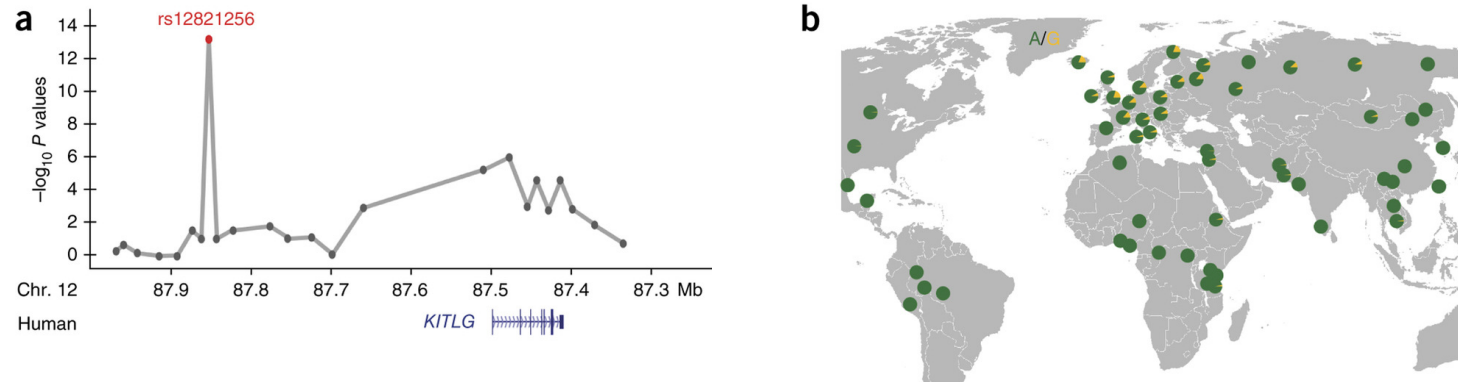
- Genetic variants linked to eight genes in humans are significantly associated with blond hair color in Europeans.
- Some variants alter the coding regions of genes known to be involved in pigmentation.
- Some variants map outside the protein-coding regions of pigmentation genes.

The Human *KITLG* gene (mouse *Kitl*)

- Encodes a secreted ligand for the KIT receptor tyrosine kinase and has an essential role in the development, differentiation and pigmentation.

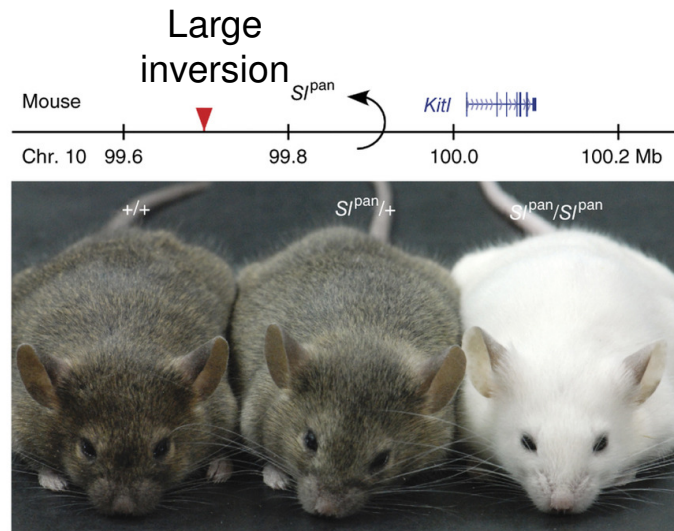


The Human *KITLG* gene (mouse *Kitl*)



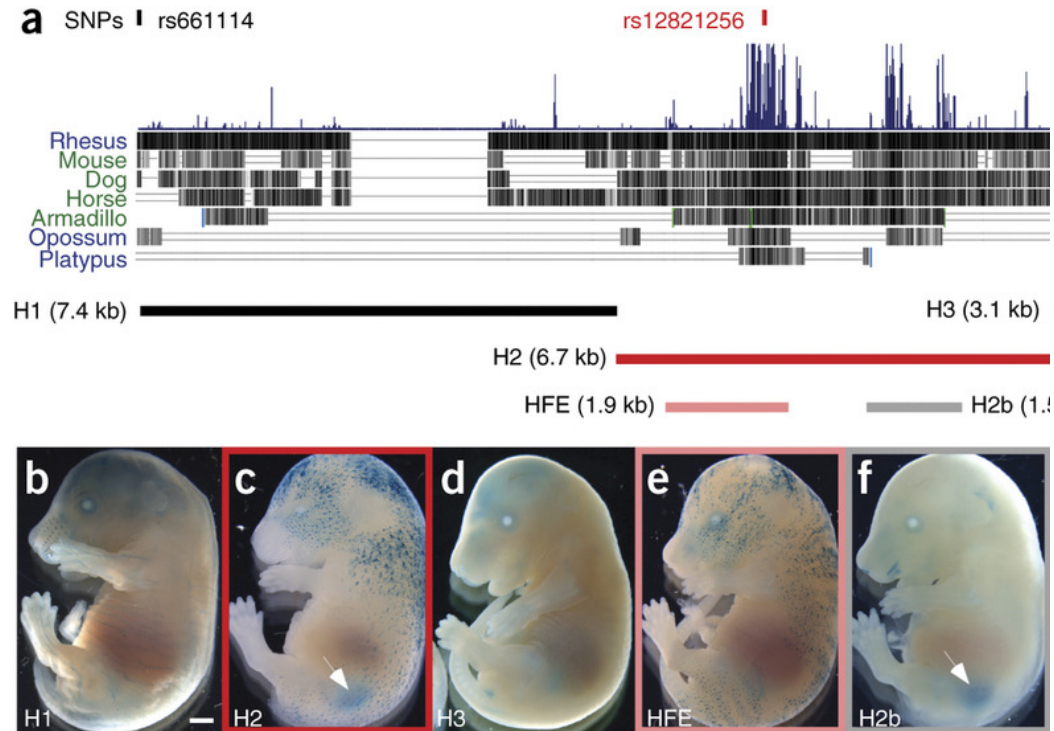
- A noncoding SNP (rs12821256) located over 350 kb upstream of *KITLG* is significantly associated with blond hair color in Iceland and The Netherlands.
- The blond-associated A>G substitution at this position is prevalent in northern European populations but virtually absent in African and Asian populations

An inversion spanning the noncoding SNP rs12821256



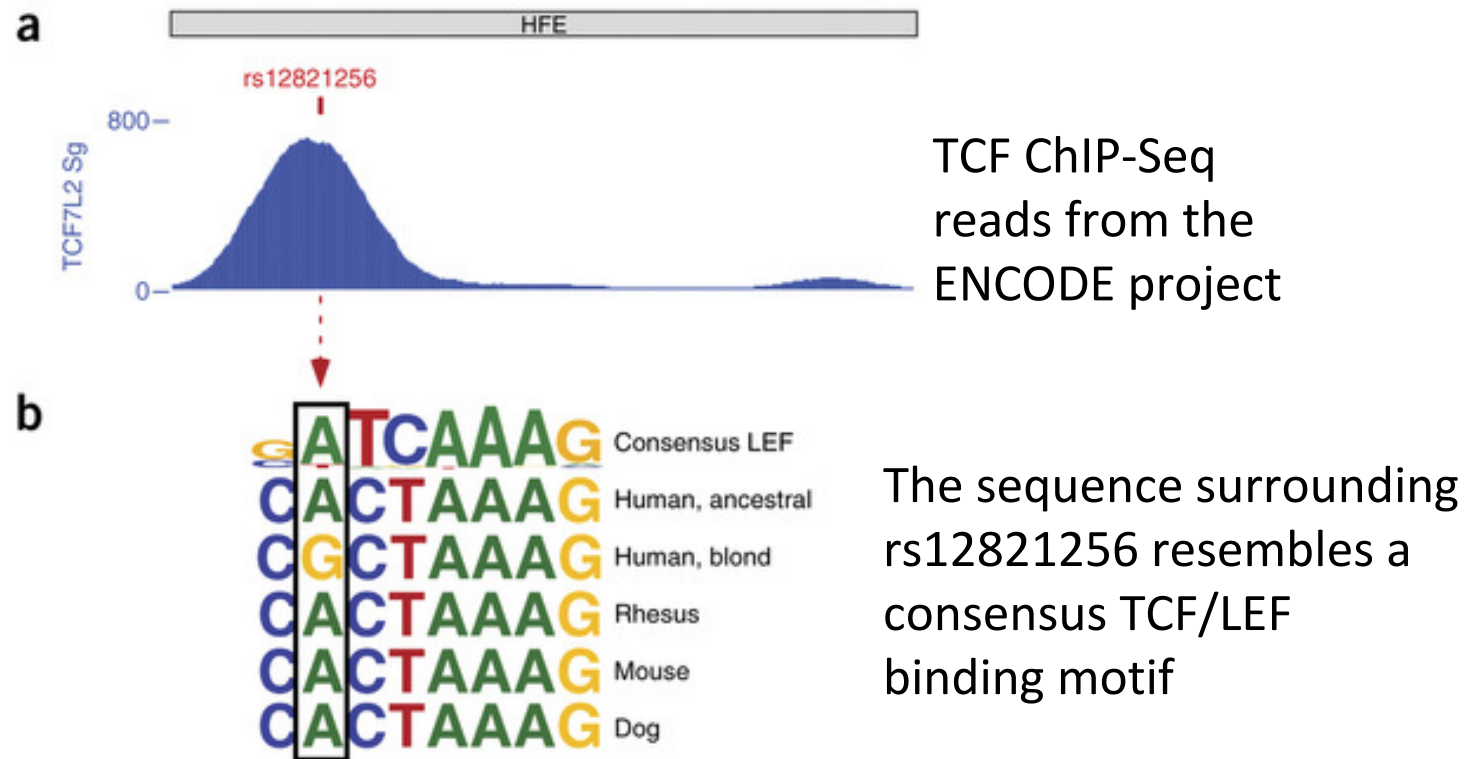
Displacement of a single copy of the distant upstream regulatory sequences for *Kitl* is sufficient to reduce *Kitl* expression and lighten hair color.

Searching for the functional enhancer

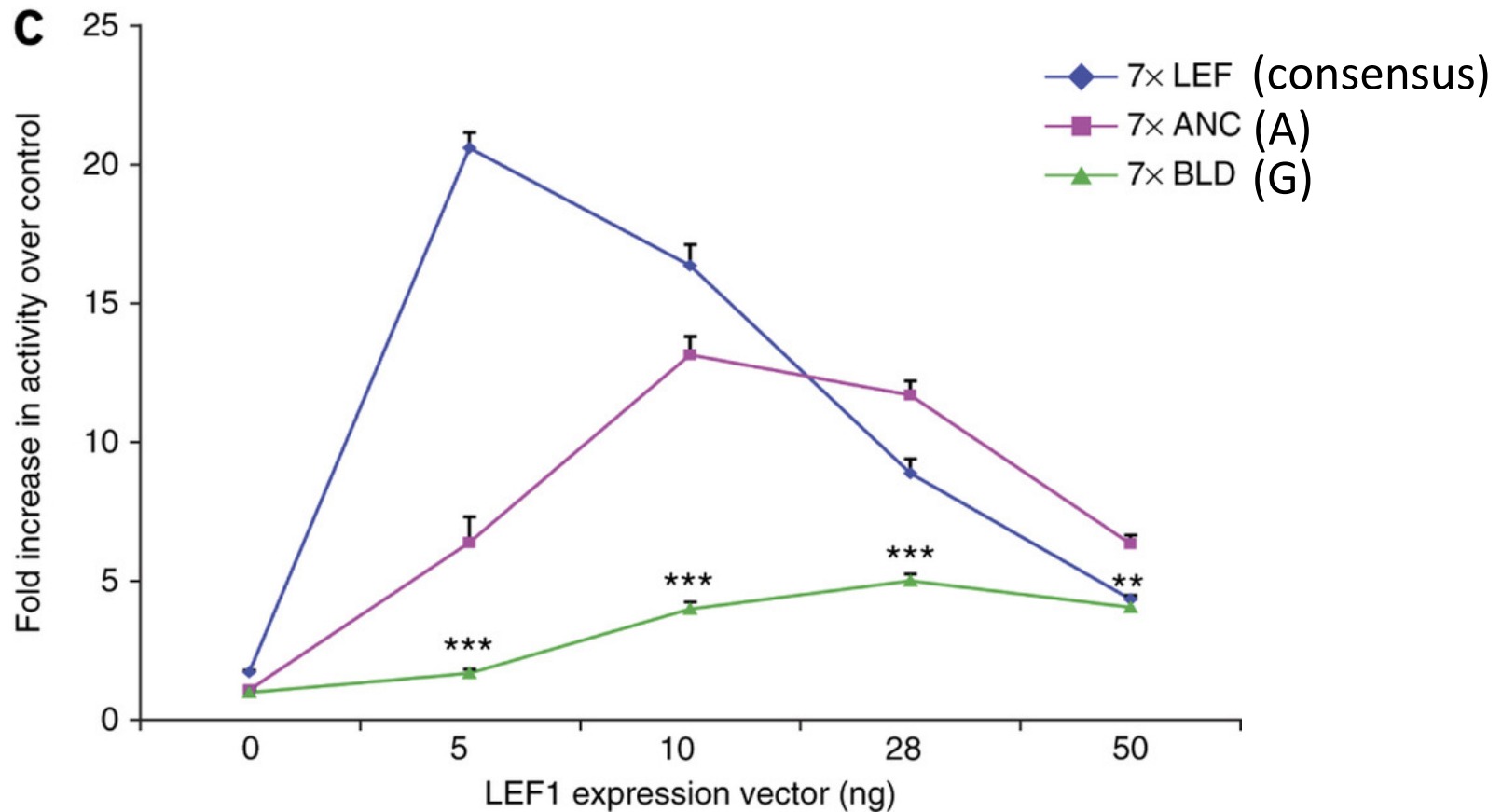


Five human fragments were cloned upstream of a *lacZ* reporter gene and tested for *in vivo* enhancer activity in transgenic mice.

rs12821256 alters a TCF/LEF binding site and reduces LEF responsiveness in keratinocytes

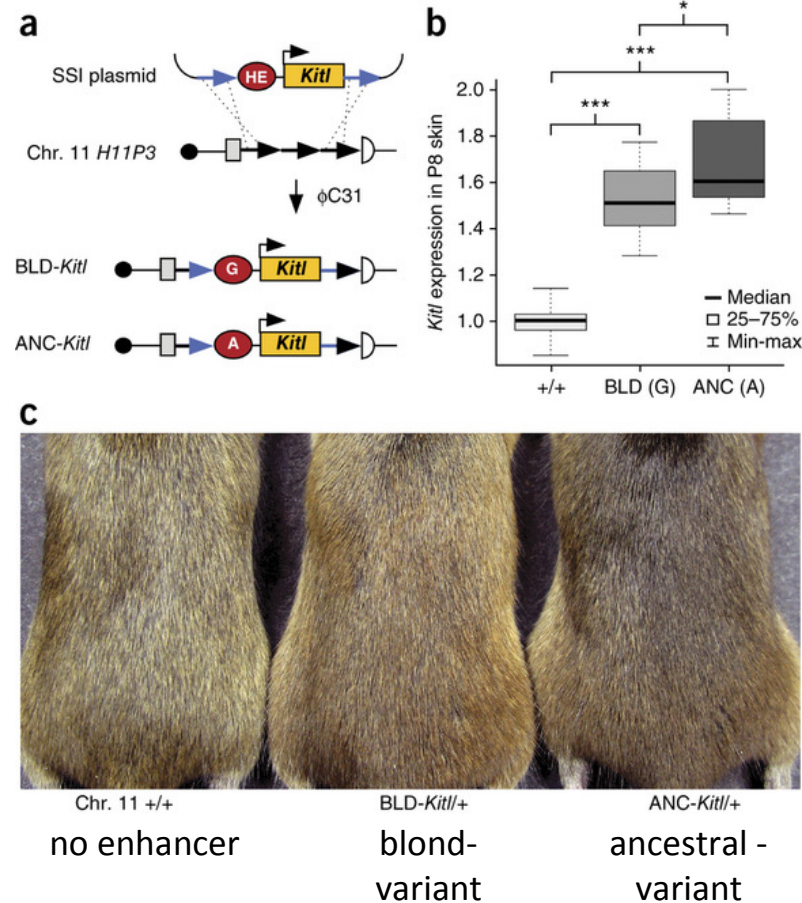


rs12821256 alters a TCF/LEF binding site and reduces LEF responsiveness in keratinocytes



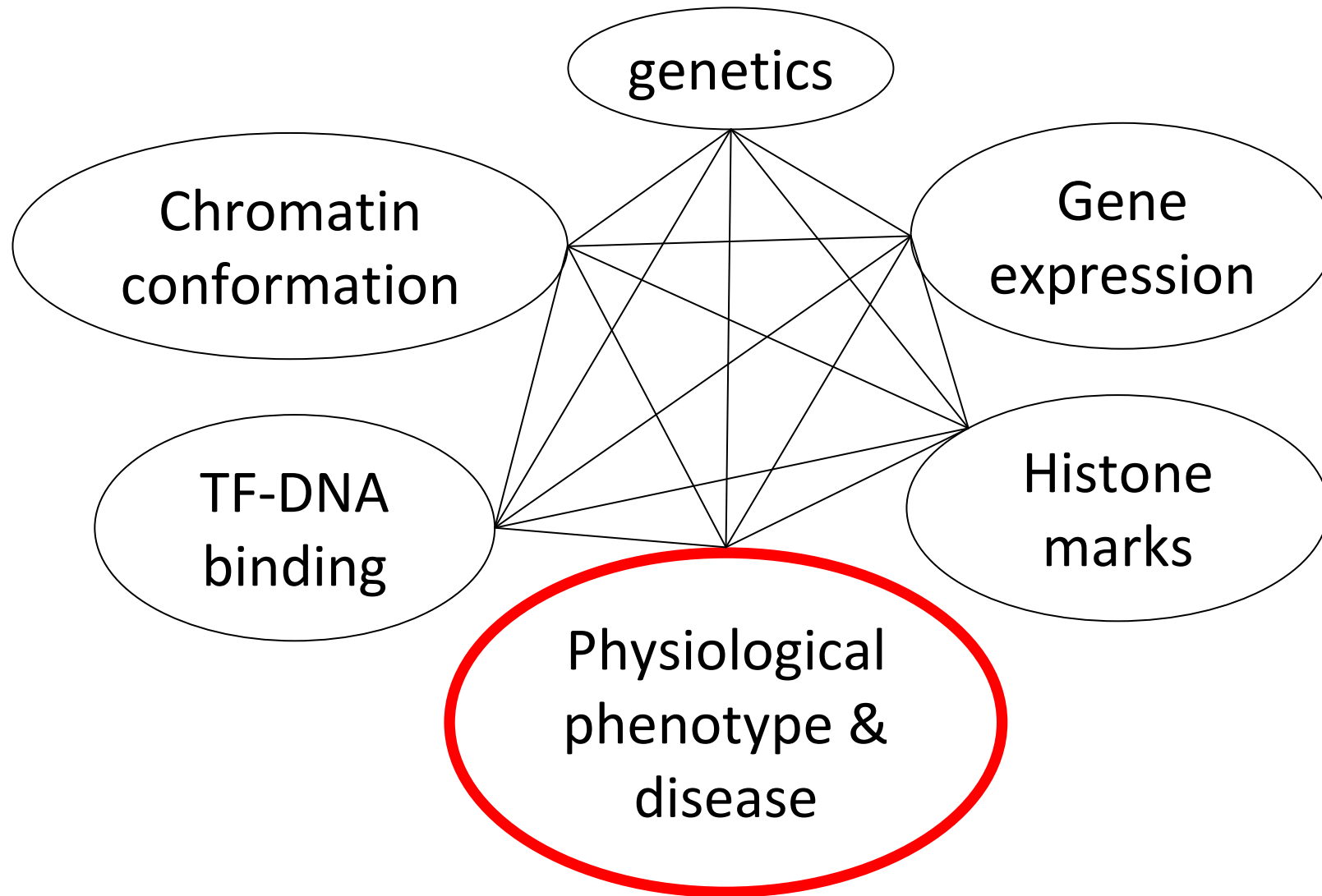
Mouse lines differing at a single base-pair position in the *KITLG* enhancer show differences in hair color

Matched lines of site-specific integration in transgenic mice



Small (20%) quantitative changes in enhancer activity were sufficient to alter hair color *in vivo*

The molecular basis of disease



Utilizing genetics to understand transcriptional circuitry and the regulatory conformation of the genome

- Interpreting susceptibility loci using epigenomic profiling
- Revealing susceptibility loci that impact the chromatin landscape: Which genetic variants determine histone marks, open chromatin and TF binding?
- Reveal functionally linked histone marks between nearby or distal regions
- Reveal chromatin regulators
- Reveal the function of chromatin states in common disease

Thank you!

